



List of Revised Courses

Department : *Biotechnology*

Program Name : *M.Sc.*

Academic Year : *2020-21*

List of Revised Courses

Sr. No.	Course Code	Name of the Course
1.	MBT 101 T	Biochemistry
2.	MBT 102T	Cell and Molecular Biology
3.	MBT 103T	Plant and Animal Biotechnology
4.	MBT 104T	Microbiology
5.	MBT 105T	Genetics
6.	MBT 106T	Biostatistics
7.	MBT 107L	Biochemistry and Analytical Techniques
8.	MBT 108L	Microbiology
9.	MBT 109L	Plant and Animal Biotechnology
10.	MBT 201 T	Genetic Engineering
11.	MBT 202T	Immunology
12.	MBT 203T	Bioinformatics
13.	MBT 204T	Genomics and Proteomics
14.	MBT 205T	Molecular Diagnostics
15.	MBT 206T	Research Methodology and Scientific Communication Skills
16.	MBT 207T	Environmental Biotechnology
17.	MBT 208T	Biological Imaging
18.	MBT 209T	Nanobiotechnology
19.	*MBT 210S	MOOCs course to be selected/opted from SWAYAM portal (SWAYAM-BIOTECH-1)
20.	MBT 211L	Molecular Biology and Genetic Engineering
21.	MBT 212 L	Immunology
22.	MBT 301 T	Bioprocess Engineering and Technology
23.	MBT 302T	Emerging Technologies



24.	MBT 303T	Critical Analysis of Classical Papers
25.	MBT 304T	Bioentrepreneurship
26.	MBT 305T	Intellectual Property Rights, Biosafety and Bioethics
27.	MBT 306T	Project Proposal Preparation and Presentation
28.	MBT 307T	Research Seminar
29.	MBT 308T	Microbial Technology
30.	MBT 309 T	Animal Biotechnology
31.	MBT 310 T	Computational Biology
32.	MBT 311 T	Drug Discovery and Development
33.	MBT 312 T	Vaccines
34.	MBT 313 T	Protein Engineering
35.	MBT 314 T	Medical Microbiology and Infection Biology
36.	*MBT 315T	MOOCs course to be selected/opted from SWAYAM portal (SWAYAM-BIOTECH-1)
37.	MBT 316L	Laboratory VI: Bioprocess Engineering and Technology
38.	MBT 317 L	Laboratory VII: Bioinformatics
39.	MBT 401	Dissertation



Minutes of Meetings (MoM) of Board of Studies (BoS)

Academic Year : 2020-21

School : School of Studies of Interdisciplinary Education and Research

Department : Biotechnology

Date and Time : 09-07-2020- 12:00 Noon

Venue : Room of Head, Department of Biotechnology

MINUTES OF THE MEETING OF BOARD OF STUDIES IN BIOTECHNOLOGY GURU GHASIDAS VISHWAVIDYALAYA, BILASPUR HELD ON 09/07/2020

A Meeting of the Board of Studies in Biotechnology under School of Interdisciplinary Education and Research was held on 09/07/2020 at 12:00 Noon under the chairmanship of Dr. Renu Bhatt, Head Department of Biotechnology. The following members were present.

(i) Dr. Renu Bhatt, Head	Chairman
(ii) Prof. B.N. Tiwary, Professor	Member
(iii) Prof. Keshavkant Sahu,	Expert present online
(iv) Dr. Dhananjay Shukla	Member

The following agenda were placed to discuss:

1. Pre Ph.D syllabus as directed by UGC (syllabus of research and publication ethics) as a compulsory first paper along with Research methodology paper I.
2. To discuss CBCS Syllabus for M.Sc programme in Biotechnology.
3. To discuss and approve the ordinance of CBCS in M.Sc Biotechnology, w.e.f. 2020-21
4. Revision of Course code of CBCS B.Sc (Hons) with revised course name IE (Interdisciplinary Education and Research) in place of LS (Life Science) w.e.f 2020-21
5. To amend and approve the credit of SEC (skill enhancement course) in as 2 instead of 4 in CBCS B.Sc (Hons) III semester as per ordinance for 2019-20.

At the very outset the HOD, Chairman of Board of Studies welcomed all the BoS members and discussed the above agenda at length. Following resolutions were made in this meeting

1. The revised Pre Ph.D course work syllabus including Research Publication Ethics in the paper I to be named as Research Methodology and Research Publication Ethics of a total of 4 credits including 2 for Research Publication Ethics as per directives of the UGC was discussed and approved by the BoS members including subject expert of BoS
2. The model syllabus of DBT (with 20% modification) CBCS M.Sc Biotechnology syllabus and scheme of a examination, the course structure with course code of 2 year M.Sc. degree course was placed before the committee. The members after a thorough deliberations approved the course structure and course code of M.Sc. Biotechnology to be implemented from the Academic session 2020-2021.
3. The draft ordinance for M.Sc Biotechnology under CBCS pattern was discussed and approved by the Board of studies and recommended to be placed before Academic Council.

R. Bhatt
9/07/2020

Dhananjay
9/7/2020

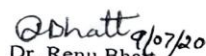
R. Singh
9/7/2020

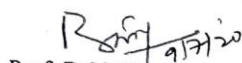


4. Since the name of School of Studies has changed from SoS of Life Science to SoS of Interdisciplinary Education and Research. The proposed revised draft of course code (LS to IE) of 3years CBCS B.Sc (Hons) was placed and approved by BoS.


5. The credit of SEC as approved by BoS for 2019-20 was discussed and resolved to be amended to 2 instead of 4 (as per existing ordinance for 2019-20).

The meeting ended with a vote of thanks by the Chairman


Dr. Renu Bhatt
Chairman


Prof. B. N. Tiwary
Member

Prof. Keshavkant Sahu
Expert present online


Dr. Dhananjay Shukla
Member

In the Meeting of BOS-Biotechnology on 09-07-2020, the following courses were revised in the syllabus of M.Sc.:

Course Code	Name of the Course
MBT 101 T	Biochemistry
MBT 102T	Cell and Molecular Biology
MBT 103T	Plant and Animal Biotechnology
MBT 104T	Microbiology
MBT 105T	Genetics
MBT 106T	Biostatistics
MBT 107L	Biochemistry and Analytical Techniques
MBT 108L	Microbiology
MBT 109L	Plant and Animal Biotechnology
MBT 201 T	Genetic Engineering
MBT 202T	Immunology
MBT 203T	Bioinformatics
MBT 204T	Genomics and Proteomics
MBT 205T	Molecular Diagnostics
MBT 206T	Research Methodology and Scientific Communication Skills
MBT 207T	Environmental Biotechnology
MBT 208T	Biological Imaging
MBT 209T	Nanobiotechnology



*MBT 210S	MOOCs course to be selected/opted from SWAYAM portal (SWAYAM-BIOTECH-1)
MBT 211L	Molecular Biology and Genetic Engineering
MBT 212 L	Immunology
MBT 301 T	Bioprocess Engineering and Technology
MBT 302T	Emerging Technologies
MBT 303T	Critical Analysis of Classical Papers
MBT 304T	Bioentrepreneurship
MBT 305T	Intellectual Property Rights, Biosafety and Bioethics
MBT 306T	Project Proposal Preparation and Presentation
MBT 307T	Research Seminar
MBT 308T	Microbial Technology
MBT 309 T	Animal Biotechnology
MBT 310 T	Computational Biology
MBT 311 T	Drug Discovery and Development
MBT 312 T	Vaccines
MBT 313 T	Protein Engineering
MBT 314 T	Medical Microbiology and Infection Biology
*MBT 315T	MOOCs course to be selected/opted from SWAYAM portal (SWAYAM-BIOTECH-1)
MBT 316L	Laboratory VI: Bioprocess Engineering and Technology
MBT 317 L	Laboratory VII: Bioinformatics
MBT 401	Dissertation

In the Meeting of BOS-Biotechnology on 09-07-2020, the following courses were revised in the of Ph.D. Course work.:

Course Code	Name of the Course
101	Research Methodology and research publication Ethics

The following new courses were introduced in the syllabus of M.Sc.:



Course Code	Name of the Course
MBT 103T	Plant and Animal Biotechnology
MBT 105T	Genetics
MBT 106T	Biostatistics
MBT 107L	Biochemistry and Analytical Techniques
MBT 109L	Plant and Animal Biotechnology
MBT 201 T	Genetic Engineering
MBT 203T	Bioinformatics
MBT 204T	Genomics and Proteomics
MBT 205T	Molecular Diagnostics
MBT 206T	Research Methodology and Scientific Communication Skills
MBT 208T	Biological Imaging
MBT 209T	Nanobiotechnology
*MBT 210S	MOOCs course to be selected/opted from SWAYAM portal (SWAYAM-BIOTECH-1)
MBT 211L	Molecular Biology and Genetic Engineering
MBT 302T	Emerging Technologies
MBT 303T	Critical Analysis of Classical Papers
MBT 305T	Intellectual Property Rights, Biosafety and Bioethics
MBT 306T	Project Proposal Preparation and Presentation
MBT 307T	Research Seminar
MBT 308T	Microbial Technology
MBT 310 T	Computational Biology
MBT 311 T	Drug Discovery and Development
MBT 312 T	Vaccines
MBT 313 T	Protein Engineering
MBT 314 T	Medical Microbiology and Infection Biology
*MBT 315T	MOOCs course to be selected/opted from SWAYAM portal (SWAYAM-BIOTECH-1)

गुरु घासीदास विश्वविद्यालय
(केन्द्रीय विश्वविद्यालय अधिनियम 2009 क्र. 25 के अंतर्गत स्थापित केन्द्रीय विश्वविद्यालय)
कोनी, बिलासपुर - 495009 (छ.ग.)



Guru Ghasidas Vishwavidyalaya
(A Central University Established by the Central Universities Act 2009 No. 25 of 2009)
Koni, Bilaspur - 495009 (C.G.)

MBT 316L	Laboratory VI: Bioprocess Engineering and Technology
MBT 317 L	Laboratory VII: Bioinformatics
MBT 401	Dissertation

Signature & Seal of HoD

विभागाध्यक्ष, जैव प्रौद्योगिकी विभाग
Head, Department of Biotechnology
गुरु घासीदास विश्वविद्यालय, बिलासपुर (छ.ग.)
Guru Ghasidas Vishwavidyalaya, Bilaspur (C.G.)



Scheme and Syllabus

Syllabus for M.Sc program in Biotechnology 2020-21

M.Sc Biotechnology PG Semester I					
code	Course opted	Subjects	Hours/ semester	Hours/ week	Credits
MBT 101T	Core -1	Biochemistry	48	03	3
MBT 102T	Core -2	Cell and Molecular Biology	48	03	3
MBT 103T	Core -3	Plant and Animal Biotechnology	48	03	3
MBT 104T	Core -4	Microbiology	32	02	2
MBT 105T	Core -5	Genetics	32	02	2
MBT 106T	Core -6	Biostatistics	48	03	3
Laboratory					
MBT 107L	Lab 01	Biochemistry and Analytical Techniques	128	08	4
MBT 108L	Lab 02	Microbiology	64	04	2
MBT 109L	Lab 03	Plant and Animal Biotechnology	64	04	2
Total			512	32	24
M.Sc Biotechnology PG Semester II					
Code	Course opted	Subjects	Hours/ semester	Hours/ week	Credits
MBT 201T	Core -1	Genetic Engineering	48	03	3
MBT 202T	Core -2	Immunology	48	03	3
MBT 203T	Core -3	Bioinformatics	48	03	3
MBT 204T	Core -4	Genomics and Proteomics	32	02	2
MBT 205T	Core -5	Molecular Diagnostics	32	02	2
MBT 206T	Core -6	Research Methodology and Scientific Communication Skills	32	02	2
MBT 207T	Elective-1	Environmental Biotechnology	32	02	2
MBT 208T	Elective-1	Biological Imaging			
MBT 209T	Elective-1	Nanobiotechnology			
*MBT 210S	Elective	MOCs course to be selected/opted from SWAYAM portal (SWAYAM-BIOTECH-1)			
Laboratory					
MBT 211L	Lab 01	Molecular Biology and Genetic Engineering	128	08	4
MBT 212L	Lab 02	Immunology	96	06	3
Total			496	31	24
M.Sc Biotechnology PG Semester III					
Code	Course opted	Subjects	Hours/ semester	Hours/ week	Credits

Abhatt 9/1/2020

Sharma 21/1/20

Prasad 10/1/20



MBT 301 T	Core -1	Bioprocess Engineering and Technology	48	03	3
MBT 302T	Core -2	Emerging Technologies	32	02	2
MBT 303T	Core -3	Critical Analysis of Classical Papers	32	02	2
MBT 304T	Core-4	Bioentrepreneurship	32	02	2
MBT 305T	Core -5	Intellectual Property Rights, Biosafety and Bioethics	32	02	2
MBT 306T	Core -6	Project Proposal Preparation and Presentation	32	02	2
MBT 307T	Core -7	Research Seminar	32	02	2
MBT 308T	Elective	Microbial Technology	48	03	3
MBT 309 T	Elective	Animal Biotechnology			
MBT 310 T	Elective	Computational Biology			
MBT 311 T	Elective	Drug Discovery and Development			
MBT 312 T	Elective	Vaccines			
MBT 313 T	Elective	Protein Engineering			
MBT 314 T	Elective	Medical Microbiology and Infection Biology			
*MBT 315T	Elective	MOOCs course to be selected/opted from SWAYAM portal (SWAYAM-BIOTECH-IT Laboratory			
MBT 316L	Lab 01	Laboratory VI: Bioprocess Engineering and Technology	128	08	4
MBT 317 L	Lab 02	Laboratory VII: Bioinformatics	64	04	2
		Total	480	30	24
M.Sc Biotechnology PG Semester IV					
Code	Course opted	Subjects	Hours/ semester	Hours/ week	Credits
MBT 401	Core -1	Dissertation	512	32	22
		Total	512	32	22
				Total	94
Credits					

*M.Sc. Biotechnology students will select Massive Open Online Course (MOOCs)-SWAYAM course in the II and III semester available at <http://ugemoocs.inflibnet.ac.in/courses.php> in consultation with Coordinator.

Chhatt
9/7/20

Sharma
9/7/20

Roy



Semester One

Biochemistry

Credits



Course Objectives

The objectives of this course are to build upon undergraduate level knowledge of biochemical principles with specific emphasis on different metabolic pathways. The course shall make the students aware of various disease pathologies within the context of each topic.

Student Learning Outcomes

On completion of this course, students should be able to:

- Gain fundamental knowledge in biochemistry;
- Understand the molecular basis of various pathological conditions from the perspective of biochemical reactions.

Unit I

Chemical basis of life and proteins 7 lectures

Chemical basis of life: **Miller-Urey experiment**, abiotic formation of amino acid oligomers, composition of living matter; Water – properties of water, **essential role of water for life on earth** pH, buffer, maintenance of blood pH and pH of gastric juice, pH optima of different enzymes (pepsin, trypsin and alkaline phosphatase), **ionization and hydrophobicity**, emergent properties of biomolecules in water, **biomolecular hierarchy**, macromolecules, molecular assemblies.

Unit II

Protein structure 4 lectures

Structure-function relationships; amino acids – structure and functional group properties, peptides and covalent structure of proteins, elucidation of primary and higher order structures, Ramachandran plot, evolution of protein structure, **protein degradation and introduction to molecular pathways controlling proteolysis**, structure-function relationships in model proteins like ribonuclease A, myoglobin, hemoglobin, chymotrypsin etc; basic principles of protein purification; tools to characterize expressed proteins; **Protein folding: Anfinsen's Dogma, Levinthal paradox, cooperativity in protein folding, free energy landscape of protein folding and pathways of protein folding**, molten globule state, chaperons, diseases associated with protein folding, introduction to molecular dynamic simulation.

Unit III

Enzyme kinetics 5 lectures

Enzyme catalysis – general principles of catalysis, quantitation of enzyme activity and efficiency; enzyme characterization and Michaelis-Menten kinetics; relevance of enzymes in metabolic regulation, activation, inhibition and covalent modification; **single substrate enzymes**; concept of catalytic antibodies; catalytic strategies with specific examples of proteases, carbonic anhydrases, restriction enzymes and nucleoside monophosphate kinase; regulatory strategies with specific example of hemoglobin; **isozymes**; role of covalent modification in enzymatic activity; zymogens.

Unit IV Glycobiology 2 lectures

Sugars - mono, di, and polysaccharides with specific reference to glycogen, **amylose and cellulose**, glycosylation of other biomolecules - glycoproteins and glycolipids; **lipids - structure and properties of important members of storage and membrane lipids**; lipoproteins.

Unit V

Structure and functions of DNA & RNA 3 lectures

Self-assembly of lipids, micelle, biomembrane organization - sidedness and function; membrane bound proteins - structure, properties and function; transport phenomena; nucleosides, nucleotides, nucleic acids - structure, a historical perspective leading up to the proposition of DNA double helical structure, difference in RNA and DNA structure and their importance in evolution of DNA as the genetic material.

Unit VI

Bioenergetics 8 lectures

Bioenergetics-basic principles; equilibria and concept of free energy; coupled interconnecting reactions in metabolism; oxidation of carbon fuels; recurring motifs in metabolism; introduction to GPCR, Inositol/DAG/PKC and Ca⁺⁺ signaling pathways;

Abstract

Post

Dr

glycolysis and gluconeogenesis; reciprocal regulations and non-carbohydrate sources of glucose; Citric acid cycle, entry to citric acid cycle, citric acid cycle as a source of biosynthetic precursors; Oxidative phosphorylation; importance of electron transfer in oxidative phosphorylation; F1-F0 ATP Synthase; shuttles across mitochondria; regulation of oxidative phosphorylation; Photosynthesis – chloroplasts and two photosystems; proton gradient across thylakoid membrane;

Unit VII

Lipids, DNA & RNA 8 lectures

Calvin cycle and pentose phosphate pathway; glycogen metabolism, reciprocal control of glycogen synthesis and breakdown, roles of epinephrine and glucagon and insulin in glycogen metabolism; Fatty acid metabolism; protein turnover and amino acid catabolism; nucleotide biosynthesis; biosynthesis of membrane lipids and sterols with specific emphasis on cholesterol metabolism and mevalonate pathway; elucidation of metabolic pathways; logic and integration of central metabolism; entry/exit of various biomolecules from central pathways; principles of metabolic regulation; steps for regulation; target of rapamycin (TOR) & Autophagy regulation in relation to C & N metabolism, starvation responses and insulin signaling. Role of vitamins & cofactors in metabolism



Recommended Textbooks and References:

1. Stryer, L. (2002). *Biochemistry*. New York: Freeman.
2. Lehninger, A. L. (2004). *Principles of Biochemistry* (4th ed.). New York, NY: Worth.
3. Voet, D., & Voet, J. G. (2004). *Biochemistry* (4th ed.). Hoboken, NJ: J. Wiley & Sons.
4. Dobson, C. M. (2003). *Protein Folding and Misfolding*. Nature, 426(6968), 884-890. doi:10.1038/nature02261.
5. Richards, F. M. (1991). *The Protein Folding Problem*. Scientific American, 264(1), 54-63. doi:10.1038/scientificamerican0191-54.



Cell and Molecular Biology

Credits



Course Objectives

The objectives of this course are to sensitize the students to the fact that as we go down the scale of magnitude from cells to organelles to molecules, the understanding of various biological processes becomes deeper and inclusive.

Student Learning Outcomes

Student should be equipped to understand three fundamental aspects in biological phenomenon: a) what to seek; b) how to seek; c) why to seek?

Unit I

Dynamic organization of cell

6 lectures

Universal features of cells; cell chemistry and biosynthesis: chemical organization of cells; internal organization of the cell - cell membranes: structure of cell membranes and concepts related to compartmentalization in eukaryotic cells; intracellular organelles: endoplasmic reticulum and Golgi apparatus, lysosomes and peroxisomes, ribosomes, cellular cytoskeleton, mitochondria, chloroplasts and cell energetics; nuclear compartment: nucleus, nucleolus and chromosomes.

Abhatt
21/7/2020

Roy
21/7/2020

Dr



Unit II
Chromatin structure and dynamics
12 lectures

Chromatin organization - histone and DNA interactions; structure and assembly of eukaryotic and prokaryotic DNA polymerases, DNA-replication, repair and recombination; chromatin control: gene transcription and silencing by chromatin-Writers,-Readers and -Erasers; Transcriptional control: Structure and assembly of eukaryotic and prokaryotic RNA Polymerases, promoters and enhancers, transcription factors as activators and repressors, transcriptional initiation, elongation and termination; post-transcriptional control: splicing and addition of cap and tail, mRNA flow through nuclear envelope into cytoplasm, breakdown of selective and specific mRNAs through interference by small non-coding RNAs (miRNAs and siRNAs), protein translation machinery, ribosomes-composition and assembly; universal genetic codes, degeneracy of codons, Wobble hypothesis; Iso-accepting tRNA; mechanism of initiation, elongation and termination; co- and post-translational modifications, mitochondrial genetic code translation product cleavage, modification and activation.

Unit III
Cellular signalling, transport and trafficking
3 lectures

Molecular mechanisms of membrane transport, nuclear transport, transport across mitochondria and chloroplasts; intracellular vesicular trafficking from endoplasmic reticulum through Golgi apparatus to lysosomes/cell exterior.

Unit IV
Cellular processes
8 lectures

Cell cycle and its regulation; cell division: mitosis, meiosis and cytokinesis; cell differentiation: stem cells, their differentiation into different cell types and organization into specialized tissues; cell-ECM and cell-cell interactions; cell receptors and trans-membrane signalling; cell motility and migration; cell death: different modes of cell death and their regulation.

Unit V
Manipulating and studying cells
3 lectures

Isolation of cells and basics of cell culture; observing cells under a microscope, different types of microscopy; analyzing and manipulating DNA, RNA and proteins.

Unit VI
Genome instability and cell transformation
8 lectures

Mutations, proto-oncogenes, oncogenes and tumour suppressor genes, physical, chemical and biological mutagens; types of mutations; intra-genic and inter-genic suppression; transpositions- transposable genetic elements in prokaryotes and eukaryotes, role of transposons in genome; viral and cellular oncogenes; tumor suppressor genes; structure, function and mechanism of action; activation and suppression of tumor suppressor genes; oncogenes as transcriptional activators.



Recommended Textbooks and References:

1. Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K., & Walter, P. (2008). *Molecular Biology of the Cell* (5th Ed.). New York: Garland Science.
2. Lodish, H. F. (2016). *Molecular Cell Biology* (8th Ed.). New York: W. H. Freeman.
3. Krebs, J. E., Lewin, B., Kilpatrick, S. T., & Goldstein, E. S. (2014). *Lewin's Genes XI*. Burlington, MA: Jones & Bartlett Learning.
4. Cooper, G. M., & Hausman, R. E. (2013). *The Cell: a Molecular Approach* (6th Ed.). Washington: ASM ; Sunderland.
5. Hardin, J., Bertoni, G., Kleinsmith, L. J., & Becker, W. M. (2012). *Becker's World of the Cell*. Boston (8th Ed.). Benjamin Cummings.
6. Watson, J. D. (2008). *Molecular Biology of the Gene* (5th ed.). Menlo Park, CA: Benjamin Cummings.

Abhishek
9/7/2020

Prof. Jaiswal

DB



Plant and Animal Biotechnology

Credits

2

Course Objectives

The objectives of this course are to introduce students to the principles, practices and application of animal biotechnology, plant tissue culture, plant and animal genomics, genetic transformation and molecular breeding of plants and animals.

Student Learning Outcomes

Students should be able to gain fundamental knowledge in animal and plant biotechnology and their applications.

Unit I Plant tissue culture and animal cell culture

10 lectures

Plant tissue culture: historical perspective; totipotency; organogenesis; Somatic embryogenesis; establishment of cultures – callus culture, cell suspension culture, media preparation – nutrients and plant hormones; sterilization techniques; applications of tissue culture - micropropagation; somaclonal variation; androgenesis and its applications in genetics and plant breeding; germplasm conservation and cryopreservation; synthetic seed production; protoplast culture and somatic hybridization - protoplast isolation; culture and usage; somatic hybridization - methods and applications; cybrids and somatic cell genetics; plant cell cultures for secondary metabolite production. Animal cell culture: brief history of animal cell culture; cell culture media and reagents; culture of mammalian cells, tissues and organs; primary culture, secondary culture, continuous cell lines, suspension cultures; application of animal cell culture for virus isolation and *in vitro* testing of drugs, testing of toxicity of environmental pollutants in cell culture, application of cell culture technology in production of human and animal viral vaccines and pharmaceutical proteins.

Unit II Plant genetic manipulation

10 lectures

Genetic engineering: Agrobacterium-plant interaction; virulence; Ti and Ri plasmids; opines and their significance; T-DNA transfer; disarmed Ti plasmid; Genetic transformation - Agrobacterium-mediated gene delivery; integrate and binary vectors and their utility; direct gene transfer - PEG-mediated, electroporation, particle bombardment and alternative methods; screenable and selectable markers; characterization of transgenics; chloroplast transformation; marker-free methodologies; advanced methodologies - cisgenesis, intragenesis and genome editing; molecular farming - concept of plants as biofactories, production of industrial enzymes and pharmaceutically important compounds.

Unit III Animal reproductive biotechnology and vaccinology

8 lectures

Animal reproductive biotechnology: structure of sperms and ovum; cryopreservation of sperms and ova of livestock; artificial insemination; super ovulation, embryo recovery and *in vitro* fertilization; culture of embryos; cryopreservation of embryos; embryo transfer technology; transgenic manipulation of animal embryos; applications of transgenic animal technology; animal cloning - basic concept, cloning for conservation for conservation endangered species; Vaccinology: history of development of vaccines, introduction to the concept of vaccines, conventional methods of animal vaccine production, recombinant approaches to vaccine production, modern vaccines.

Unit IV Plant and animal genomics

4 lectures

Overview of genomics – definition, complexity and classification; need for genomics level analysis; methods of analyzing genome at various levels – DNA, RNA, protein, metabolites and phenotype; genome projects and bioinformatics resources for genome research – databases; overview of forward and reverse genetics for assigning function for genes.

08/04/2020

08/04/2020

08/04/2020



Unit V
Molecular mapping and marker assisted selection
8 lectures

Molecular markers - hybridization and PCR based markers RFLP, RAPD, STS, SSR, AFLP, SNP markers; DNA fingerprinting-principles and applications; introduction to mapping of genes/QTLs; marker-assisted selection - strategies for introducing genes of biotic and abiotic stress resistance in plants; genetic basis for disease resistance in animals; molecular diagnostics of pathogens in plants and animals; detection of meat adulteration using DNA based methods.



Recommended Textbooks and References:

1. Charila, H. S. (2006). *Introduction to Plant Biotechnology*. Enfield, NH: Science.
2. Razdan, M. K. (2003). *Introduction to Plant Tissue Culture*. Enfield, NH: Science.
3. Slater, A., Scott, N. W. & Fowler, M. R. (2008). *Plant Biotechnology: an Introduction to Genetic Engineering*. Oxford: Oxford University Press.
4. Buchanan, B. B., Gruissem, W., & Jones, R. L. (2015). *Biochemistry & Molecular Biology of Plants*. Chichester, West Sussex: John Wiley & Sons.
5. Umesh, S. (2013). *Plant Biotechnology: The Energy And Resources*.
6. Glick, B. R., & Pasternak, J. J. (2010). *Molecular Biotechnology: Principles and Applications of Recombinant DNA*. Washington, D.C.: ASM Press.
7. Brown, T. A. (2006). *Gene Cloning and DNA Analysis: an Introduction*. Oxford: Blackwell Pub.
8. Primrose, S. B., & Twyman, R. M. (2006). *Principles of Gene Manipulation and Genomics*. Malden, MA: Blackwell Pub.
9. Slater, A., Scott, N. W., & Fowler, M. R. (2003). *Plant Biotechnology: The Genetic Manipulation of Plants*. Oxford: Oxford University Press.
10. Gordon, I. (2005). *Reproductive Techniques in Farm Animals*. Oxford: CAB International.
11. Levine, M. M. (2004). *New Generation Vaccines*. New York: M. Dekker.
12. Pfitzer, R. (2007). *Animal Cell Biotechnology: Methods and Protocols*. Totowa, NJ: Humana Press.

Microbiology

Credits

2

Course Objectives

The objectives of this course are to introduce field of microbiology with special emphasis on microbial diversity, morphology, physiology and nutrition; methods for control of microbes and host-microbe interactions.

Student Learning Outcomes

- Students should be able to:
- Identify major categories of microorganisms and analyze their classification, diversity, and ubiquity;
 - Identify and demonstrate structural, physiological, genetic similarities and differences of major categories of microorganisms;
 - Identify and demonstrate how to control microbial growth;
 - Demonstrate and evaluate interactions between microbes, hosts and environment.

Unit I
Microbial characteristics
6 lectures

Introduction to microbiology and microbes, history & scope of microbiology, morphology, structure, growth and nutrition of bacteria, bacterial growth curve, bacterial culture methods; bacterial genetics: mutation and recombination in bacteria, plasmids, transformation, transduction and conjugation; antimicrobial resistance.

Unit II
Microbial diversity
9 lectures

Microbial taxonomy and evolution of diversity, classification of microorganisms, criteria for classification; classification of bacteria: Cyanobacteria, acetic acid bacteria, Pseudomonads, lactic and propionic acid bacteria, endospore forming bacteria.

Abhijit 21/7/2020

Rohit 21/7/20

AB



Unit III Control of microorganisms 3 lectures	Mycobacteria and Mycoplasma, Archaea: Halophiles, Methanogens, Hyperthermophilic archaea, Thermophiles; eukarya: algae, fungi, slime molds and protozoa; extremophiles and unculturable microbes.
Unit IV Virology 5 lectures	Virus and bacteriophages, general properties of viruses, viral structure, taxonomy of virus, viral replication, cultivation and identification of viruses; sub-viral particles – viroids and prions.
Unit V Host-microbes interaction 5 lectures	Host-pathogen interaction, ecological impact of microbes; symbiosis (Nitrogen fixation and ruminant symbiosis); microbes and nutrient cycles; microbial communication system; bacterial quorum sensing; microbial fuel cells; probiotics and prebiotics.



Recommended Textbooks and References:

1. Pelczar, M. J., Reid, R. D., & Chan, E. C. (2001). Microbiology (5th ed.). New York: McGraw-Hill.
2. Willey, J. M., Sherwood, L., Woolverton, C. J., Prescott, L. M., & Willey, J. M. (2011). Prescott's Microbiology. New York: McGraw-Hill.
3. Mathai, W., Berg, C. V., & Black, J. G. (2005). Microbiology, Principles and Explorations. Boston, MA: John Wiley & Sons.

Genetics

Credits

2

Course Objectives

The objectives of this course are to take students through basics of genetics and classical genetics covering prokaryotic/ phage genetics to yeast and higher eukaryotic domains. On covering all classical concepts of Mendelian genetics across these life-forms, students will be exposed to concepts of population genetics, quantitative genetics encompassing complex traits, clinical genetics and genetics of evolution.

Student Learning Outcomes

On successful completion of this course, student will be able to:

- Describe fundamental molecular principles of genetics;
- Understand relationship between phenotype and genotype in human genetic traits;
- Describe the basics of genetic mapping;
- Understand how gene expression is regulated.

Unit I
Genetics of bacteria and bacteriophages
10 lectures

Concept of a gene in pre-DNA era; mapping of genes in bacterial and phage chromosomes by classical genetic crosses; fine structure analysis of a gene; genetic complementation and other genetic crosses using phenotypic markers; phenotype to genotype connectivity prior to DNA-based understanding of gene.

Unit II
Yeast genetics
8 lectures

Meiotic crosses, tetrad analyses, non-Mendelian and Mendelian ratios, gene conversion, models of genetic recombination, yeast mating type switch; dominant and recessive gene mutations, suppressor or modifier screens, complementation groups, transposon mutagenesis, synthetic lethality, genetic epistasis.

Obhett
9/1/2020

Rohit

DL



Unit III
Drosophila genetics as a model of higher eukaryotes
4 lectures

Monohybrid & dihybrid crosses, back-crosses, test-crosses, analyses of autosomal and sex linkages, screening of mutations based on phenotypes and mapping the same, hypomeophy, genetic mosaics, genetic epistasis in context of developmental mechanism.

Unit IV
Population genetics and genetics of evolution
4 lectures

Introduction to the elements of population genetics: genetic variation, genetic drift, sexual evolution; mutation selection, balancing selection, Fisher theorem, Hardy-Weinberg equilibrium, linkage disequilibrium; in-breeding depression & mating systems; population bottlenecks, migrations, Bayesian statistics; adaptive landscape, spatial variation & genetic fitness.

Unit V
Quantitative genetics of complex traits (QTLs)
2 lectures

Complex traits, mapping QTLs, yeast genomics to understand biology of QTLs.

Unit VI
Plant genetics
2 lectures

Laws of segregation in plant crosses, inbreeding, selfing, heterosis, maintenance of genetic purity, gene pyramiding.



Recommended Textbooks and References:

- Hartl, D. L., & Jones, E. W. (1998). *Genetics: Principles and Analysis*. Sudbury, MA: Jones and Bartlett.
- Pierce, B. A. (2005). *Genetics: a Conceptual Approach*. New York: W.H. Freeman.
- Tamarin, R. H., & Leavitt, R. W. (1991). *Principles of Genetics*. Dubuque, IA: Wm. C. Brown.
- Smith, J.M. (1998). *Evolutionary Genetics*. Oxford: Oxford University Press.

Bio-Statistics

Credits

2

Course Objectives

The objective of this course is to give conceptual exposure of statistics, error analysis, hypothesis testing, and design of experiments in biological systems

Student Learning Outcomes

On completion of this course, students should be able to:

- Understand how to summarise statistical data;
- Apply appropriate statistical tests based on an understanding of study question, type of study and type of data;
- Interpret results of statistical tests and application in biological systems.

Unit I
Introduction
5 lectures

Types of biological data (ordinal scale, nominal scale, continuous and discrete logical systems data), frequency distribution and graphical representations (bar graph, histogram, box plot and frequency polygon), cumulative frequency distribution, populations, samples, simple random, stratified and systematic sampling.

Abhijit
9/7/2020

Rajesh

03



Unit II Descriptive statistics, Probability and distribution 10 lectures	Measures of Location, Properties of Arithmetic Mean, median, mode, range, Properties of the Variance and Standard Deviation, Coefficient of Variation, Grouped Data, Graphic Methods, Obtaining Descriptive Statistics on the Computer, Case study, Introduction to probability and laws of probability, Random Events, Events-exhaustive, Mutually exclusive and equally likely (with simple exercises), Definition and properties of binomial distribution, Poisson distribution and normal distribution.
Unit III Correlation and regression analysis, Statistical hypothesis 10 lectures	Correlation, Covariance, calculation of covariance and correlation, Correlation coefficient from ungrouped data, Spearson's Rank Correlation Coefficient, scatter and dot diagram, General Concepts of regression, Fitting Regression Lines, regression coefficient, properties of Regression Coefficients, Standard error of estimate, Making assumption, Null and alternate hypothesis, error in hypothesis testing, confidence interval, one-tailed and two-tailed testing, decision making. Making assumption, Null and alternate hypothesis, error in hypothesis testing, confidence interval, one-tailed and two-tailed testing, decision making.
Unit IV Tests of significance 8 lectures	Steps in testing statistical significance, selection and computation of test of significance and interpretation of results, Sampling distribution of mean and standard error, Large sample tests (test for an assumed mean and equality of two population means with known S.D.), z-test; Small sample tests (t-test for an assumed mean and equality of means of two populations when sample observations are independent); parametric and Non parametric tests (Mann-Whitney test); paired and unpaired t-test, chi square test.
Unit V Experimental designs 8 lectures	Introduction to study designs: Longitudinal, cross-sectional, retrospective and prospective study, Principles of experimental designs, Randomized block, and Simple factorial designs, Analysis of variance (ANOVA) and its use in analysis of RBD, introduction to meta-analysis and systematic reviews, ethics in statistics.



Recommended Textbooks and References:

1. Jaype Brothers, (2011), Methods in Biostatistics for Medical Students and Research Workers (English), 7th Edition
2. Norman T.J. Bailey, (1995), Statistical Methods in Biology, 3rd Edition, Cambridge University Press.
3. P. N. Arora and P. K. Malhan, (2006), Biostatistics, 2nd Edition, Himalaya Publishing House.
4. Jerold Zar, Biostatistical Analysis, 4th Edition, Pearson Education.
5. Biostatistics: a Foundation for Analysis in the Health Sciences, 7th Edition, Wiley.
6. ML Samuels, JA Witmer (2003) Statistics for the Life Sciences, 3rd edition. Prentice Hall.

Abhakt
4/7/2020

Raj 4/7/2020

Dr



Laboratory I: Biochemistry & Analytical Techniques

Credits



Course Objectives

The objective of this laboratory course is to introduce students to experiments in biochemistry. The course is designed to teach students the utility of set of experimental methods in biochemistry in a problem oriented manner.

Student Learning Outcomes

On completion of this course, students should be able to:

- To elaborate concepts of biochemistry with easy to run experiments;
- To familiarize with basic laboratory instruments and understand the principle of measurements using those instruments with experiments in biochemistry.

- Preparing various stock solutions and working solutions that will be needed for the course.
- To prepare an Acetic-Na Acetate Buffer and validate the Henderson-Hasselbalch equation.
- To determine an unknown protein concentration by plotting a standard graph of BSA using UV-Vis Spectrophotometer and validating the Beer-Lambert's Law.
- Titration of Amino Acids and separation of aliphatic, aromatic and polar amino acids by thin layer chromatography.
- Purification and characterization of an enzyme from a recombinant source (such as Alkaline Phosphatase or Lactate Dehydrogenase or any enzyme of the institution's choice).
 - Preparation of cell-free lysates
 - Ammonium Sulfate precipitation
 - Ion-exchange Chromatography
 - Gel Filtration
 - Affinity Chromatography
 - Dialysis of the purified protein solution against 60% glycerol as a demonstration of storage method
 - Generating a Purification Table (protein concentration, amount of total protein; Computing specific activity of the enzyme preparation at each stage of purification)
 - Assessing purity of samples from each step of purification by SDS-PAGE Gel Electrophoresis
 - Enzyme Kinetic Parameters: Km, Vmax and Kcat.
- Experimental verification that absorption at OD₂₆₀ is more for denatured DNA as compared to native double stranded DNA, reversal of the same following DNA renaturation. Kinetics of DNA renaturation as a function of DNA size.
- Identification of an unknown sample as DNA, RNA or protein using available laboratory tools. (Optional Experiments)
- Biophysical methods (Circular Dichroism Spectroscopy, Fluorescence Spectroscopy).
- Determination of mass of small molecules and fragmentation patterns by Mass Spectrometry.
- Preparing various stock solutions and working solutions that will be needed for the course.

Dr. Anshu
21/7/2020

Ravi
21/7/2020

Dr.



Laboratory II: Microbiology

Credits



Course Objectives

The objective of this laboratory course is to provide practical skills on basic microbiological techniques.

Student Learning Outcomes

Students should be able to:

- Isolate, characterize and identify common bacterial organisms;
- Determine bacterial load of different samples;
- Perform antimicrobial sensitivity tests;
- Preserve bacterial cultures.

Syllabus

1. Sterilization, disinfection and safety in microbiological laboratory.
2. Preparation of media for cultivation of bacteria.
3. Isolation of bacteria in pure culture by streak plate method.
4. Study of colony and growth characteristics of some common bacteria:
Bacillus, E. coli, Staphylococcus, Streptococcus, etc.
5. Preparation of bacterial smear and Gram's staining.
6. Enumeration of bacteria: standard plate count.
7. Antimicrobial sensitivity test and demonstration of drug resistance.
8. Maintenance of stock cultures: slants, stabs and glycerol stock cultures
9. Determination of phenol co-efficient of antimicrobial agents.
10. Determination of Minimum Inhibitory Concentration (MIC)
11. Isolation and identification of bacteria from soil/water samples.



Recommended Textbooks and References:

1. Cappuccino, J.G., & Welsh, C. (2016). *Microbiology: a Laboratory Manual*. Benjamin-Cummings Publishing Company.
2. Collins, C. H., Lyne, P.M., Grange, J. M., & Falkinham III, J. (2004). *Collins and Lyne's Microbiological Methods* (8th ed.). Arnold.
3. Tille, P.M., & Forbes, B. A. *Bailey & Scott's Diagnostic Microbiology*.



Laboratory III: Plant and Animal Biotechnology

Credits



Course Objectives

The objectives of this course are to provide hands-on training in basic experiments of plant and animal biotechnology.

Student Learning Outcomes

On completion of course, students should be able to gain basic skills in plant and animal biotechnology.

Syllabus

Plant Biotechnology

1. Prepare culture media with various supplements for plant tissue culture.
2. Prepare explants of *Vallerianawallichii* for inoculation under aseptic conditions.
3. Attempt *in vitro* andro and gyno genesis in plants (*Daturastramonium*).
4. Isolate plant protoplast by enzymatic and mechanical methods and attempt fusion by PEG (available material).
5. Culture *Agrobacteriumtumefaciens* and attempt transformation of any dicot species.
6. Generate RAPD and SSR profile of *Eremuruspersicus* and *Vallerianawallichii*.
7. Prepare karyotypes and study the morphology of somatic chromosomes of *Allium cepa*, *A.sativum*, *A.tuberosum* and compare them on the basis of karyotypes.
8. Pollen mother cell meiosis and recombination index of selected species (one achiasmate, and the other chiasmate) and correlate with generation of variation.
9. Undertake plant genomic DNA isolation by CTAB method and its quantitation by visual as well as spectrophotometric methods.
10. Perform PCR amplification of 'n' number of genotypes of a species for studying the genetic variation among the individuals of a species using random primers.
11. Study genetic fingerprinting profiles of plants and calculate polymorphic information content.

Syllabus

Animal Biotechnology

1. Count cells of an animal tissue and check their viability.
2. Prepare culture media with various supplements for plant and animal tissue culture.
3. Prepare single cell suspension from spleen and thymus.
4. Monitor and measure doubling time of animal cells.
5. Chromosome preparations from cultured animal cells.
6. Isolate DNA from animal tissue by SDS method.
7. Attempt animal cell fusion using PEG.



Semester Two

Genetic Engineering

Credits
3

Course Objectives

The objectives of this course are to teach students with various approaches to conducting genetic engineering and their applications in biological research as well as in biotechnology industries. Genetic engineering is a technology that has been developed based on our fundamental understanding of the principles of molecular biology and this is reflected in the contents of this course.

Student Learning Outcomes

Given the impact of genetic engineering in modern society, the students should be endowed with strong theoretical knowledge of this technology. In conjunction with the practicals in molecular biology & genetic engineering, the students should be able to take up biological research as well as placement in the relevant biotech industry.

**Unit I
Introduction and tools for genetic engineering**
6 lectures

Impact of genetic engineering in modern society; general requirements for performing a genetic engineering experiment; restriction endonucleases and methylases; DNA ligase; Klenow enzyme, T4 DNA polymerase, polynucleotide kinase, alkaline phosphatase; cohesive and blunt end ligation; linkers; adaptors; homopolymeric tailing; labelling of DNA; nick translation, random priming, radioactive and non-radioactive probes, hybridization techniques: northern, southern, south-western and far-western and colony hybridization, fluorescence in situ hybridization.

**Unit II
Different types of vectors**
7 lectures

Plasmids; Bacteriophages; M13 mp vectors; PUC19 and Bluescript vectors, hogenids; Lambda vectors; Insertion and Replacement vectors; Cosmids; Artificial chromosome vectors (YACs; BACs); Principles for maximizing gene expression expression vectors; pMal; GST; pET-based vectors; Protein purification; His-tag; GST-tag; MBP-tag etc; Intron-based vectors; Inclusion bodies; methodologies to reduce formation of inclusion bodies; mammalian expression and replicating vectors; Baculovirus and Pichia vectors system, plant based vectors, Ti and Ri as vectors, yeast vectors, shuttle vectors.

**Unit III
Different types of PCR techniques**
7 lectures

Principles of PCR: primer design; fidelity of thermostable enzymes; DNA polymerases; types of PCR – multiplex, nested, reverse-transcription PCR, real time PCR, touchdown PCR, hot start PCR, colony PCR, asymmetric PCR, cloning of PCR products; T-vectors; proof reading enzymes; PCR based site specific mutagenesis; PCR in molecular diagnostics; viral and bacterial detection; sequencing methods; enzymatic DNA sequencing; chemical sequencing of DNA; automated DNA sequencing; RNA sequencing; chemical synthesis of oligonucleotides; mutation detection: SSCP, DGGE, RFLP.

**Unit IV
Gene manipulation and protein-DNA interaction**
7 lectures

Insertion of foreign DNA into host cells; transformation, electroporation, transfection; construction of libraries; isolation of mRNA and total RNA; reverse transcriptase and cDNA synthesis; cDNA and genomic libraries; construction of microarrays – genomic arrays, cDNA arrays and oligo arrays; study of protein-DNA interactions; electrophoretic mobility shift assay; DNase footprinting; methyl interference assay, chromatin immunoprecipitation; protein-protein interactions using yeast two-hybrid system; phage display.

**Unit V
Gene silencing and genome editing technologies**
13 lectures

Gene silencing techniques; introduction to siRNA, siRNA technology; Micro RNA; construction of siRNA vectors; principle and application of gene silencing; gene knockouts and gene therapy; creation of transgenic plants; debate over GM crops; introduction to methods of genetic manipulation in different model systems e.g. fruit flies

*Okhett
21/7/2020*

Prof. 21/7/2020

Pr



(Drosophila), worms (*C. elegans*), frogs (*Xenopus*), fish (zebra fish) and chick; Transgenesis - gene replacement; gene targeting; creation of transgenic and knock-out mice; disease model; introduction to genome editing by CRISPR-CAS with specific emphasis on Chinese and American clinical trials.



Recommended Textbooks and References:

1. Old, R. W., Primrose, S. B., & Twayman, R. M. (2001). *Principles of Gene Manipulation: an Introduction to Genetic Engineering*. Oxford: Blackwell Scientific Publications.
2. Green, M. R., & Sambrook, J. (2012). *Molecular Cloning: a Laboratory Manual*. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press.
3. Brown, T. A. (2006). *Genomes* (3rd ed.). New York: Garland Science Pub.
4. Selected papers from scientific journals, particularly Nature & Science.
5. Technical Literature from Stratagene, Promega, Novagen, New England Biolab etc.

Immunology

Credits

3

Course Objectives

The objectives of this course are to learn about structural features of components of immune system as well as their function. The major emphasis of this course will be on development of immune system and mechanisms by which our body elicits immune response. This will be imperative for students as it will help them to predict about nature of immune response that develops against bacterial, viral or parasitic infection, and prove it by designing new experiments.

Student Learning Outcomes

On completion of this course, students should be able to:

- Evaluate usefulness of immunology in different pharmaceutical companies;
- Identify proper research lab working in area of their own interests;
- Apply their knowledge and design immunological experiments to demonstrate innate, humoral or cytotoxic T lymphocyte responses and figure out kind of immune responses in the setting of infection (viral or bacterial).

Unit I

Immunology: fundamental concepts and overview of the immune system
5 lectures

Components of innate and acquired immunity; phagocytosis; complement and inflammatory responses; pathogen recognition receptors (PRR) and pathogen associated molecular pattern (PAMP); innate immune response; mucosal immunity; antigens: immunogens, haptens; Major Histocompatibility Complex: MHC genes, MHC and immune responsiveness and disease susceptibility. Organs of immune system, primary and secondary lymphoid organs.

Unit II

Immune responses generated by B and T lymphocytes
8 lectures

Immunoglobulins - basic structure, classes & subclasses of immunoglobulins, antigenic determinants, multigene organization of immunoglobulin genes; B-cell receptor; Immunoglobulin superfamily; principles of cell signaling; basis of self & non-self discrimination; kinetics of immune response, memory; B cell maturation, activation and differentiation; generation of antibody diversity; T-cell maturation, activation and differentiation and T-cell receptors; functional T Cell subsets; cell-mediated immune responses, ADCC; cytokines: properties, receptors and therapeutic uses; antigen processing and presentation- endogenous antigens, exogenous antigens, non-peptide bacterial antigens and super-antigens; cell-cell co-operation, Hapten-carrier system.

Unit III

Antigen-antibody interactions
6 lectures

Precipitation, agglutination and complement mediated immune reactions; advanced immunological techniques: RIA, ELISA, Western blotting, ELISPOT assay, immunofluorescence microscopy, flow cytometry and immunoelectron microscopy; surface plasmon resonance, biosensor assays for assessing ligand-receptor interaction; CMI techniques: lymphoproliferation assay, mixed lymphocyte reaction, cell cytotoxicity assays, apoptosis, microarrays, transgenic mice, gene knock out.

Obtained 21/7/2022

Prof. Anshu

Dr.



Unit IV

Vaccinology
8 lectures

Active and passive immunization; live, killed, attenuated, subunit vaccines; vaccine technology; role and properties of adjuvants, recombinant DNA and protein based vaccines, plant-based vaccines, reverse vaccinology; peptide vaccines, conjugate vaccines; antibody genes and antibody engineering; chimeric, generation of monoclonal antibodies, hybrid monoclonal antibodies; catalytic antibodies and generation of immunoglobulin gene libraries, idiotype vaccines and marker vaccines, viral-like particles (VLPs), dendritic cell based vaccines, vaccine against cancer, T cell based vaccine, edible vaccine and therapeutic vaccine.

Unit V

Clinical immunology
8 lectures

Immunity to infection : bacteria, viral, fungal and parasitic infections (with examples from each group); hypersensitivity: Type I-IV; autoimmunity: types of autoimmune diseases; mechanism and role of CD4+ T cells; MHC and TCR in autoimmunity; treatment of autoimmune diseases; transplantation: immunological basis of graft rejection; clinical transplantation and immunosuppressive therapy; tumor immunology: tumor antigens; immune response to tumours and tumor evasion of the immune system, cancer immunotherapy; immunodeficiency: primary immunodeficiencies, acquired or secondary immunodeficiencies, autoimmune disorder, anaphylactic shock, immunosenescence, immune exhaustion in chronic viral infection, immune tolerance, NK cells in chronic viral infection and malignancy.

Unit VI

Immunogenetics
5 lectures

Major histocompatibility complex genes and their role in autoimmune and infectious diseases, HLA typing, human major histocompatibility complex (MHC), Complement genes of the human major histocompatibility complex: implication for linkage disequilibrium and disease associations, genetic studies of rheumatoid arthritis, systemic lupus erythematosus and multiple sclerosis, genetics of human immunoglobulin, immunogenetics of spontaneous control of HIV, KIR complex.



Recommended Textbooks and References:

1. Kiehl, T. J., Goldby, R. A., Osborne, B. A., & Kuby, J. (2006). *Kuby Immunology*. New York: W. H. Freeman.
2. Brastoff, J., Seaddin, J. K., Male, D., & Roitt, I. M. (2002). *Clinical Immunology*. London: Gower Medical Pub.
3. Murphy, K., Travers, P., Walport, M., & Janeway, C. (2012). *Janeway's Immunobiology*. New York: Garland Science.
4. Paul, W. E. (2012). *Fundamental Immunology*. New York: Raven Press.
5. Goding, J. W. (1996). *Monoclonal Antibodies: Principles and Practice: Production and Application of Monoclonal Antibodies in Cell Biology, Biochemistry, and Immunology*. London: Academic Press.
6. Parham, P. (2005). *The Immune System*. New York: Garland Science.

Bioinformatics

Credits

3

Course Objectives

The objectives of this course are to provide theory and practical experience of the use of common computational tools and databases which facilitate investigation of molecular biology and evolution-related concepts.

Student Learning Outcomes

Student should be able to :

- Develop an understanding of basic theory of these computational tools;
- Gain working knowledge of these computational tools and methods;
- Appreciate their relevance for investigating specific contemporary biological questions;
- Critically analyse and interpret results of their study.

Q. D. White

R. Singh

Q. D. White



<p>Unit I Bioinformatics basics 5 lectures</p>	<p>Bioinformatics basics: Computers in biology and medicine; Introduction to Unix and Linux systems and basic commands; Database concepts; Proteins and nucleic acid databases; Structural databases; Biological XML DTD's; pattern matching algorithm basics; databases and search tools; biological background for sequence analysis; Identification of protein sequence from DNA sequence; searching of databases similar sequence; NCBI; publicly available tools; resources at EBI; resources on web; database mining tools.</p>
<p>Unit II DNA sequence analysis 5 lectures</p>	<p>DNA sequence analysis: gene bank sequence database; submitting DNA sequences to databases and database searching; sequence alignment; pairwise alignment techniques; motif discovery and gene prediction; local structural variants of DNA, their relevance in molecular level processes, and their identification; assembly of data from genome sequencing.</p>
<p>Unit III Multiple sequence analysis 5 lectures</p>	<p>Multiple sequence analysis; multiple sequence alignment; flexible sequence similarity searching with the FASTA3 program package; use of CLUSTALW and CLUSTALX for multiple sequence alignment; submitting DNA protein sequence to databases: where and how to submit, SEQUIN, genome centres; submitting aligned sets of sequences, updating submitted sequences, methods of phylogenetic analysis.</p>
<p>Unit IV Protein modelling 5 lectures</p>	<p>Protein modelling: introduction; force field methods; energy, buried and exposed residues; side chains and neighbours; fixed regions; hydrogen bonds; mapping properties onto surfaces; fitting monomers; RMS fit of conformers; assigning secondary structures; sequence alignment- methods, evaluation, scoring; protein completion; backbone construction and side chain addition; small peptide methodology; software accessibility; building peptides; protein displays; substructure manipulations; annealing.</p>
<p>Unit V Protein structure prediction and virtual library 6 lectures</p>	<p>Protein structure prediction: protein folding and model generation; secondary structure prediction; analyzing secondary structures; protein loop searching; loop generating methods; homology modelling: potential applications, description, methodology; homologous sequence identification; align structures, align model sequence; construction of variable and conserved regions; threading techniques; topology fingerprint approach for prediction; evaluation of alternate models; structure prediction on a mystery sequence; structure aided sequence techniques of structure prediction; structural profiles, alignment algorithms, mutation tables, prediction, validation, sequence based methods of structure prediction, prediction using inverse folding, fold prediction; significance analysis, scoring techniques, sequence-sequence scoring; protein function prediction; elements of in silico drug design; Virtual library: Searching PubMed, current content, science citation index and current awareness services, electronic journals, grants and funding information.</p>



Recommended Textbooks and References:

1. Lesk, A. M. (2002). *Introduction to Bioinformatics*. Oxford: Oxford University Press.
2. Mount, D. W. (2001). *Bioinformatics: Sequence and Genome Analysis*. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press.
3. Bavevanis, A. D., & Ouellette, B. F. (2001). *Bioinformatics: a Practical Guide to the Analysis of Genes and Proteins*. New York: Wiley-Interscience.
4. Pevsner, J. (2015). *Bioinformatics and Functional Genomics*. Hoboken, NJ.: Wiley-Blackwell.
5. Bourne, P.E., & Gu, J. (2009). *Structural Bioinformatics*. Hoboken, NJ: Wiley-Liss.
6. Lesk, A.M. (2004). *Introduction to Protein Science: Architecture, Function, and Genomics*. Oxford: Oxford University Press.

Obhatt

Booth

OB



Genomics and Proteomics

Credits

2

Course Objectives

The objectives of this course is to provide introductory knowledge concerning genomics, proteomics and their applications.

Student Learning Outcomes

Students should be able to acquire knowledge and understanding of fundamentals of genomics and proteomics, transcriptomics and metabolomics and their applications in various applied areas of biology.

Unit I Basics of genomics and proteomics 3 lectures	Brief overview of prokaryotic and eukaryotic genome organization; extra-chromosomal DNA; bacterial plasmids, mitochondria and chloroplast.
Unit II Genome mapping 4 lectures	Genetic and physical maps; markers for genetic mapping; methods and techniques used for gene mapping, physical mapping, linkage analysis, cytogenetic techniques, FISH technique in gene mapping, somatic cell hybridization, radiation hybrid maps, in situ hybridization, comparative gene mapping.
Unit III Genome sequencing projects 3 lectures	Human Genome Project, genome sequencing projects for microbes, plants and animals, accessing and retrieving genome project information from the web.
Unit IV Comparative genomics 5 lectures	Identification and classification of organisms using molecular markers- 16S (rRNA) typing/sequencing, SNPs; use of genomes to understand evolution of eukaryotes, track emerging diseases and design new drugs; determining gene location in genome sequence.
Unit V Proteomics 5 lectures	Aims, strategies and challenges in proteomics; proteomics technologies: 2D-PAGE, isoelectric focusing, mass spectrometry, MALDI-TOF, yeast 2-hybrid system, proteome databases.
Unit VI Functional genomics and proteomics 8 lectures	Transcriptome analysis for identification and functional annotation of gene, Contig assembly, chromosome walking and characterization of chromosomes, mining functional genes in genome, gene function- forward and reverse genetics, gene ethics; protein-protein and protein-DNA interactions; protein chips and functional proteomics; clinical and biomedical applications of proteomics; introduction to metabolomics, lipidomics, metagenomics and systems biology.



Recommended Textbooks and References:

1. Primrose, S. B., Twyman, R. M., Primrose, S. B., & Primrose, S. B. (2006). *Principles of Gene Manipulation and Genomics*. Malden, MA: Blackwell Pub.
2. Liebler, D. C. (2002). *Introduction to Proteomics: Tools for the New Biology*. Totowa, NJ: Humana Press.
3. Campbell, A. M., & Hoyer, L. J. (2001). *Discovering Genomics, Proteomics, and Bioinformatics*. San Francisco: Benjamin Cummings.

Abhatt

Ranjana

B3



Molecular Diagnostics

Credits

2

Course Objectives

The objectives of this course are to sensitize students about recent advances in molecular biology and various facets of molecular medicine which has potential to profoundly alter many aspects of modern medicine including pre- or post-natal analysis of genetic diseases and identification of individuals predisposed to disease ranging from common cold to cancer.

Student Learning Outcomes

Students should be able to understand various facets of molecular procedures and basics of genomics, proteomics and metabolomics that could be employed in early diagnosis and prognosis of human diseases.

Unit I Genome biology in health and disease 4 lectures

DNA, RNA, Protein: An overview; chromosomal structure & mutations; DNA polymorphism; human identity; clinical variability and genetically determined adverse reactions to drugs.

Unit II Genome: resolution, detection & analysis 5 lectures

PCR: Real-time; ARMS; Multiplex; ISH; FISH; ISA; RFLP; DHPLC; DGGE; CSCE; SSCP; Nucleic acid sequencing; new generations of automated sequencers; Microarray chips; EST; SAGE; microarray data normalization & analysis; molecular markers: 16S rRNA typing; Diagnostic proteomics; SELDI-TOF-MS; Bioinformatics data acquisition & analysis.

Unit III Diagnostic metabolomics 2 lectures

Metabolic profile for biomarker detection the body fluids/tissues in various metabolic disorders by making using LCMS & NMR technological platform.

Unit IV Detection and identity of microbial diseases 4 lectures

Direct detection and identification of pathogenic organisms that are slow growing or currently lacking a system of *in vitro* cultivation as well as genotypic markers of microbial resistance to specific antibiotics.

Unit V Detection of inherited diseases 4 lectures

Exemplified by two inherited diseases for which molecular diagnosis has provided a dramatic improvement of quality of medical care: Fragile X Syndrome: Paradigm of new mutational mechanism of unstable triplet repeats, von-Hippel Lindau disease: recent acquisition in growing number of familial cancers syndromes.

Unit VI Molecular oncology 5 lectures

Detection of recognized genetic aberrations in clinical samples from cancer patients; types of cancer-causing alterations revealed by next-generation sequencing of clinical isolates; predictive biomarkers for personalized onco-therapy of human diseases such as chronic myeloid leukemia, colon, breast, lung cancer and melanoma as well as matching targeted therapies with patients and preventing toxicity of standard systemic therapies.

Unit VII Quality assurance and control 1 lecture

Quality oversight; regulations and approved testing.



Recommended Textbooks and References:

1. Campbell, A. M., & Hoyer, L. J. (2006). *Discovering Genomics, Proteomics, and Bioinformatics*. San Francisco: Benjamin Cummings.
2. Brooker, R. J. (2009). *Genetics: Analysis & Principles*. New York, NY: McGraw-Hill.



Research Methodology and Scientific Communication Skills

Credits

2

Course Objectives

The objectives of this course are to give background on history of science, emphasizing methodologies used to do research, use framework of these methodologies for understanding effective lab practices and scientific communication and appreciate scientific ethics.

Student Learning Outcomes

Students should be able to:

- 1. Understand history and methodologies of scientific research, applying these to recent published papers;
- 2. Understand and practice scientific reading, writing and presentations;
- 3. Appreciate scientific ethics through case studies.

Unit I History of science and science methodologies 8 lectures

Empirical science; scientific method; manipulative experiments and controls; deductive and inductive reasoning; descriptive science; reductionist vs holistic biology.

Unit II Preparation for research 2 lectures

Choosing a mentor, lab and research question; maintaining a lab notebook.

Unit III Process of communication 5 lectures

Concept of effective communication- setting clear goals for communication; determining outcomes and results; initiating communication; avoiding breakdowns while communicating; creating value in conversation; barriers to effective communication; non-verbal communication- interpreting non-verbal cues; importance of body language; power of effective listening; recognizing cultural differences; Presentation skills - formal presentation skills; preparing and presenting using over-head projector, PowerPoint; defending interrogation; scientific poster preparation & presentation; participating in group discussions; Computing skills for scientific research - web browsing for information search; search engines and their mechanism of searching; hidden Web and its importance in scientific research; internet as a medium of interaction between scientists; effective email strategy using the right tone and conciseness.

Unit IV Scientific communication 9 lectures

Technical writing skills - types of reports; layout of a formal report; scientific writing skills - importance of communicating science; problems while writing a scientific document; plagiarism, software for plagiarism; scientific publication writing; elements of a scientific paper including abstract, introduction, materials & methods, results, discussion, references; drafting titles and framing abstracts; publishing scientific papers- peer review process and problems, recent developments such as open access and non-blind review; plagiarism; characteristics of effective technical communication; scientific presentations; ethical issues; scientific misconduct.

Abhatt

Raj

M



Laboratory IV: Molecular Biology and Genetic Engineering

Credits



Course Objectives

The objectives of this course are to provide students with experimental knowledge of molecular biology and genetic engineering.

Student Learning Outcomes

Students should be able to gain hands-on experience in gene cloning, protein expression and purification. This experience would enable them to begin a career in industry that engages in genetic engineering as well as in research laboratories conducting fundamental research.

Syllabus

1. Concept of lac-operon
 - a) Lactose induction of β -galactosidase.
 - b) Glucose Repression.
 - c) Diauxic growth curve of *E.coli*
2. UV mutagenesis to isolate amino acid auxotroph
3. Phage titre with epsilon phage/M13
4. Genetic Transfer-Conjugation, gene mapping
5. Plasmid DNA isolation and DNA quantitation
6. Restriction Enzyme digestion of plasmid DNA
7. Agarose gel electrophoresis
8. Polymerase Chain Reaction and analysis by agarose gel electrophoresis
9. Vector and Insert Ligation
10. Preparation of competent cells
11. Transformation of *E.coli* with standard plasmids, Calculation of transformation efficiency
12. Confirmation of the insert by Colony-PCR and Restriction mapping
13. Expression of recombinant protein, concept of soluble proteins and inclusion body formation in *E.coli*, SDS-PAGE analysis
14. Purification of His-Tagged protein on Ni-NTA columns
 - a) Random Primer labeling
 - b) Southern hybridization.



Recommended Textbooks and References:

1. Green, M. R., & Sambrook, J. (2012). *Molecular Cloning: a Laboratory Manual*. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press.

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Laboratory V: Immunology

Credits



Course Objectives

The objectives of this laboratory course are to develop an understanding about practical aspects of components of immune system as well as their function. Basic as well as advanced methods will be taught to detect different antigen and antibody interactions, isolation of different lymphocyte cells etc. and how they can be used in respective research work.

Student Learning Outcomes

Students should be able to:

- Evaluate usefulness of immunology in different pharmaceutical companies;
- Identify proper research lab working in area of their own interests;
- Apply their knowledge and design immunological experiments to demonstrate innate, humoral or cytotoxic T lymphocyte responses and figure out kind of immune response in setting of infection (viral or bacterial) by looking at cytokine profile.

Syllabus

1. Selection of animals, preparation of antigens, immunization and methods of blood collection, serum separation and storage.
2. Antibody titre by ELISA method.
3. Double diffusion, Immuno-electrophoresis and Radial Immuno diffusion.
4. Complement fixation test.
5. Isolation and purification of IgG from serum or IgY from chicken egg.
6. SDS-PAGE, Immunoblotting, Dot blot assays.
7. Blood smear identification of leucocytes by Giemsa stain.
8. Separation of leucocytes by dextran method.
9. Demonstration of Phagocytosis of latex beads and their cryopreservation.
10. Separation of mononuclear cells by Ficoll-Hypaque and their cryopreservation.
11. Demonstration of ELISPOT.
12. Demonstration of FACS.

Semester Three

Bioprocess Engineering & Technology

Credits



Course Objectives

The objectives of this course are to educate students about the fundamental concepts of bioprocess technology and its related applications, thus preparing them to meet the challenges of the new and emerging areas of biotechnology industry.

Student Learning Outcomes

Students should be able to:

- Appreciate relevance of microorganisms from industrial context;
- Carry out stoichiometric calculations and specify models of their growth;
- Give an account of design and operations of various fermenters;
- Present unit operations together with the fundamental principles for basic methods in production technique for bio-based products;
- Calculate yield and production rates in a biological production process, and also interpret data;
- Calculate the need for oxygen and oxygen transfer;
- Critically analyze any bioprocess from market point of view;
- Give an account of important microbial/enzymatic industrial processes in food and fuel industry.

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Unit I Basic principles of biochemical engineering 4 lectures	Isolation, screening and maintenance of industrially important microbes; microbial growth and death kinetics (an example from each group, particularly with reference to industrially useful microorganisms); strain improvement for increased yield and other desirable characteristics.
Unit II Stoichiometry and models of microbial growth 4 lectures	Elemental balance equations; metabolic coupling – ATP and NAD ⁺ ; yield coefficients; unstructured models of microbial growth; structured models of microbial growth.
Unit III Bioreactor design and analysis 8 lectures	Batch and continuous fermenters; modifying batch and continuous reactors: chemostat with recycle, multistage chemostat systems, fed-batch operations; conventional fermentation vs biotransformation; immobilized cell systems; large scale animal and plant cell cultivation; fermentation economics; upstream processing: media formulation and optimization; sterilization; aeration, agitation and heat transfer in bioprocess; scale up and scale down; measurement and control of bioprocess parameters.
Unit IV Downstream processing and product recovery 8 lectures	Separation of insoluble products - filtration, centrifugation, sedimentation, flocculation; Cell disruption; separation of soluble products: liquid-liquid extraction, precipitation, chromatographic techniques, reverse osmosis, ultra and micro filtration, electrophoresis; final purification: drying, crystallization; storage and packaging.
Unit V Fermentation economics 4 lectures	Isolation of micro-organisms of potential industrial interest; strain improvement; market analysis; equipment and plant costs; media; sterilization, heating and cooling; aeration and agitation; batch-process cycle times and continuous cultures; recovery costs; water usage and recycling; effluent treatment and disposal.
Unit VI Applications of enzyme technology in food processing 4 lectures	Mechanism of enzyme function and reactions in process techniques; enzymatic bioconversions e.g. starch and sugar conversion processes; high-fructose corn syrup; interesterified fat; hydrolyzed protein etc. and their downstream processing; baking by amylases, deoxygenation and desogaring by glucosyl oxidase, beer mashing and chill proofing; cheese making by proteases and various other enzyme catalytic actions in food processing.
Unit VII Applications of microbial technology in food process operations and production, biofuels and biorefinery 4 lectures	Fermented foods and beverages; food ingredients and additives prepared by fermentation and their purification; fermentation as a method of preparing and preserving foods; microbes and their use in pickling, producing colour and flavours, alcoholic beverages and other products; process wastes-whey, molasses, starch substrates and other food wastes for bioconversion to useful products; bacteriocins from lactic acid bacteria – production and applications in food preservation; biofuels and biorefinery



Recommended Textbooks and References:

1. Staler, M. L., & Kargi, F. (2002). *Bioprocess Engineering: Basic Concepts*. Upper Saddle River, NJ: Prentice Hall.
2. Stanbury, P.F., & Whitaker, A. (2010). *Principles of Fermentation Technology*. Oxford: Pergamon Press.
3. Blanch, H. W., & Clark, D.S. (1997). *Biochemical Engineering*. New York: M. Dekker.
4. Bailey, J. E., & Ollis, D. F. (1986). *Biochemical Engineering Fundamentals*. New York: McGraw-Hill.

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Emerging Technologies

Credits

2

Course Objectives

This course is broad-based in nature encompassing several new technologies that current experimental researchers are employing to probe complex systems biology questions in life-sciences. The objectives of this course are to teach basics of the new principles to students so as to appreciate current-day research tool-kit better.

Student Learning Outcomes

Students should be able to learn history, theoretical basis and basic understanding of latest technologies in area of biotechnology. They should also be able to learn about various applications of these technologies. The student may also learn one application in depth through an assignment and/or seminar.

Unit I Optical microscopy methods 6 lectures

Basic Microscopy: Light Microscopy: lenses and microscopes, resolution; Rayleigh's Approach, Darkfield; Phase Contrast; Differential Interference Contrast; fluorescence and fluorescence microscopy: what is fluorescence, what makes a molecule fluorescent, fluorescence microscope; optical arrangement, light source; filter sets: excitation filter, dichroic mirror, and barrier, optical layout for image capture; CCD camera; back illumination, binning; recording color; three CCD elements with dichroic beamsplitters, boosting the signal.

Unit II Mass spectroscopy 4 lectures

ionization techniques; mass analyzers/overview MS; FT-ICR and Orbitrap, fragmentation of peptides; proteomics, nanoLC-MS; Phosphoproteomics; interaction proteomics; mass spectroscopy in structural biology; imaging mass spectrometry.

Unit III Systems biology 3 lectures

High throughput screens in cellular systems, target identification, validation of experimental methods to generate the omics data, bioinformatics analyses, mathematical modeling and designing testable predictions.

Unit IV Structural biology 3 lectures

X-ray diffraction methods, solution & solid-state NMR, cryo-electron microscopy, small-angle X-ray scattering, Atomic force microscopy.

Unit V CRISPR-CAS 6 lectures

History of its discovery, elucidation of the mechanism including introduction to all the molecular players, development of applications for its use in genome engineering for genetic studies, promise of the technology as a next generation therapeutic method.

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Unit VI
Nanobodies
4 lectures

Introduction to nanobodies, combining nanobody with phage-display method for development of antibody against native proteins, nanobody as a tool for protein structure-function studies, use of nanobodies for molecular imaging, catalytic antibodies using nanobodies.



Recommended Textbooks and References:

1. Campbell, I. D. (2012). *Biophysical Techniques*. Oxford: Oxford University Press.
2. Serdyuk, I. N., Zaccari, N. R., & Zaccari, G. (2007). *Methods in Molecular Biophysics: Structure, Dynamics, Function*. Cambridge: Cambridge University Press.
3. Phillips, R., Kondev, J., & Theriot, J. (2009). *Physical Biology of the Cell*. New York: Garland Science.
4. Nelson, P. C., Radesavlevic, M., & Bromberg, S. (2004). *Biological Physics: Energy, Information, Life*. New York: W.H. Freeman.
5. Huang, B., Bates, M., & Zhuang, X. (2009). *Super-Resolution Fluorescence Microscopy*. *Annual Review of Biochemistry*, 78(1), 993-1016. doi:10.1146/annurev-biochem.77.061906.092014.
6. Mohamjud, P., Mikarova, K. S., Zetsche, B., Zhang, F., Koonin, E. V., & Cost, J. V. (2016). *Diverse Evolutionary Roots and Mechanistic Variations of the CRISPR-Cas Systems*. *Science*, 353(6299). doi:10.1126/science.125147.
7. Lander, E. (2016). *The Heroes of CRISPR*. *Cell*, 164(1-2), 13-28. doi:10.1016/j.cell.2015.12.041.
8. Ledford, H. (2016). *The Unseen Heroes of CRISPR*. *Nature*, 535(7612), 342-344. doi:10.1038/535342a.
9. Jinek, M., Chylinski, K., Fonfara, I., Hauer, M., Doudna, J. A., & Charpentier, E. (2012). *A Programmable Dual-RNA-Guided DNA Endonuclease in Adaptive Bacterial Immunity*. *Science*, 337(6096), 816-821. doi:10.1126/science.1225829.
10. Hamers-Casterman, C., Atarouch, T., Muylderma, S., Robinson, G., Hamers, C., Songa, E. B., Hamers, R. (1993). *Naturally Occurring Antibodies Devoid of Light Chains*. *Nature*, 363(6428), 446-448. doi:10.1038/363446a0.
11. Sidhu, S. S., & Koide, S. (2007). *Phage Display for Engineering and Analyzing Protein Interaction Interfaces*. *Current Opinion in Structural Biology*, 17(4), 481-487. doi:10.1016/j.sbi.2007.08.007.
12. Sieyaert, J., & Kobilka, B. K. (2011). *Nanobody Stabilization of G Protein-Coupled Receptor Conformational States*. *Current Opinion in Structural Biology*, 21(4), 567-572. doi:10.1016/j.sbi.2011.06.011.
13. Vincke, C., & Muylderma, S. (2012). *Introductory to Heavy Chain Antibodies and Derived Nanobodies*. *Single Domain Antibodies*, 15-26. doi:10.1007/978-1-61779-968-6_2.
14. Verheesen, P., & Laereans, T. (2012). *Selection by Phage Display of Single Domain Antibodies Specific to Antigens in their Native Conformation*. *Single Domain Antibodies*, 81-104. doi:10.1007/978-1-61779-968-6_6.
15. Li, J., Xia, L., Su, Y., Liu, H., Xia, X., Lu, Q., Reheman, K. (2012). *Molecular Imprint of Enzyme Active Site by Camel Nanobodies*. *Journal of Biological Chemistry*. *J. Biol. Chem.*, 287(17), 13713-13721. doi:10.1074/jbc.m111.336370.
16. Sohler, J., Laurent, C., Chevignat, A., Pardon, E., Srinivasan, V., Wernery, U., Galleni, M. (2013). *Allosteric Inhibition of VIM Metallo- β -Lactamases by a Camelid Nanobody*. *Biochemical Journal*, 450(3), 477-486. doi:10.1042/bj20121305.
17. Chakravarty, R., Goel, S., & Cai, W. (2014). *Nanobody: The "Magic Bullet" for Molecular Imaging?* *Theranostics*, 4(4), 386-398. doi:10.7150/thns.8006.

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Critical Analysis of Classical Papers

Credits

2

Course Objectives

The objectives of this course are to familiarize students with classic literature to make them appreciate how ground-breaking discoveries were made without, necessarily, use of high-end technologies.

Student Learning Outcomes

Students should be able to train in the exercise of hypothesis building and methods of addressing the hypothesis with readily available technology.

How does the Course Module work? Students may be divided in groups and each group may be responsible for one classical paper. Each week there may be a 1.5 hour presentation cum discussion for each of the papers. At the end of the semester each student will be asked to write a mini-review (2-3 pages long) on any one classical paper, other than the one he/she presented/discussed.

A list of sixteen classic papers and some suggested reference materials:

Syllabus

Molecular Biology

1. Studies on the chemical nature of the substance inducing transformation of Pneumococcal types: Induction of transformation by a desoxyribonucleic acid fraction isolated from *Pneumococcus* type III.
Avery OT, Macleod CM, McCarty M.; *J Exp Med.* 1944 Feb 1;79(2):137-58.
Note: This paper demonstrates that DNA is the transforming Principle originally described by Fredrick Griffith.
2. Independent functions of viral protein and nucleic acid in growth of bacteriophage
Hershey AD and Chase M.; *J Gen Physiol.* 1952 May;36(1):39-56.
Note: Note: This paper demonstrates that DNA, and not protein, component of phages enter bacterial cells.
3. Molecular structure of nucleic acids; a structure for deoxyribose nucleic acid
Watson JD and Crick FH; *Nature.* 1953 Apr 25;171(4356):737-8
Note: In this one page paper Watson and Crick first described the structure of DNA double helix.
Study help - Watson_Crick_Nature_1953_annotated
4. Transposable mating type genes in *Saccharomyces cerevisiae*
James Hicks, Jeffrey N. Strathern & Amar J.S. Klar; *Nature* 282, 478-483, 1979
Note: This paper provided evidence for 'cassette hypothesis' of yeast mating type switches i.e. interconversion of mating types in yeast (*S. cerevisiae*) occurs by DNA rearrangement.
5. Meselson & Stahl experiment demonstrating semi-conservative replication of DNA.
Meselson M and Stahl FW.; *Proc Natl Acad Sci U S A.* 1958 Jul 15;44(7):671-82.
Note: The experiment demonstrating semi-conservative mode of DNA replication is referred to as "the most beautiful experiment in biology"
6. In vivo alteration of telomere sequences and senescence caused by mutated Tetrahymena telomerase RNAs
Guo-Liang Ye, John D. Bradley, Laura D. Attardi & Elizabeth H. Blackburn; *Nature* 344, 126-132, 1990
Note: This paper demonstrates that the telomerase contains the template for telomere synthesis

Syllabus

Cell Biology

1. A protein-conducting channel in the endoplasmic reticulum
Simon SM AND Blobel G.; *Cell.* 1991 May 3;65(3):371-80
Note: This paper demonstrates the existence of a protein conducting channel
Study help - A brief history of Signal Hypothesis

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Prof. Jyoti

B



2. Identification of 23 complementation groups required for post-translational events in the yeast secretory pathway
Novick P, Field C, Schekman R.; Cell. 1980 Aug;21(1):205-15
Note: In this groundbreaking paper Randy Schekman's group used a mutagenesis screen for fast sedimenting yeast mutants to identify genes involved in cell secretion
3. A yeast mutant defective at an early stage in import of secretory protein precursors into the endoplasmic reticulum
Deshaies RJ and Schekman R.; J Cell Biol. 1987 Aug;105(2):633-45
Note: Using another yeast mutation screen Schekman lab identifies Sec61, a component of ER protein Conducting Channel (PCC)
Suggested reference paper - A biochemical assay for identification of PCC.
4. Reconstitution of the Transport of Protein between Successive Compartments of the Golgi
Balch WE, Duphy WG, Braell WA, Rothman JE.; Cell. 1984 Dec;39(2 Pt 1):405-16
Note: This paper describes setting up of an *in vitro* reconstituted system for transport between golgi stacks which eventually paved the way for identification of most of the molecular players involved in these steps including NSF, SNAP etc.
5. A complete immunoglobulin gene is created by somatic recombination
Brack C, Hinana M, Lenhard-Schuller R, Tonegawa S.; Cell. 1978 Sep;15(1):1-14
Note: This study demonstrates DNA level molecular details of somatic rearrangement of immunoglobulin gene sequences leading to the generation of functionally competent antibody generating gene following recombination.
6. A novel multigene family may encode odorant receptors: a molecular basis for odor recognition
Buck L and Axel R.; Cell. 1991 Apr 5;65(1):175-87
Note: This paper suggests that different chemical odorants associate with different cell-specific expression of a transmembrane receptor in *Drosophila* olfactory epithelium where a large family of odorant receptors is expressed.
7. Kinesin walks hand-over-hand
Yildiz A, Tomishige M, Vale RD, Selvin PR.; Science. 2004 Jan 30;303(5658):676-8
Note: This paper shows that kinesin motor works as a two-headed dimeric motor walking hand-over-hand rather than like an inchworm on microtubule tract using the energy of ATP hydrolysis.

Syllabus
Developmental
Biology/ Genetics

1. Mutations affecting segment number and polarity in *Drosophila*
Christiane Nüsslein-Volhard and Eric Wieschaus; Nature 387, 795-801, 1980
Note: This single mutagenesis screen identified majority of the developmentally important genes not only in flies but in other metazoans as well.
2. Information for the dorsal-ventral pattern of the *Drosophila* embryo is stored as maternal mRNA
Anderson KV and Nüsslein-Volhard C; Nature. 1984 Sep 20;311(5983):223-7
Note: This landmark paper demonstrated that early dorsal-ventral pattern information is stored as maternal mRNA in flies and devised the method of identifying genes encoding such genes
3. Hedgehog signalling in the mouse requires intralagellar transport proteins
Huangfu D, Liu A, Rakeman AS, Murcia NS, Niswander L, Anderson KV; Nature. 2003 Nov 6;426(6962):83-7
Note: One of the architects of original fly mutagenesis screens conducted a mouse mutagenesis screen which identified a gene KIF3a as a major component of hedgehog signaling pathway. Eventually this discovery revolutionizes our understanding of mechanisms of action of signaling pathways by demonstrating central role of cilia in it.
Suggested Reference paper - Design and execution of a embryonic lethal mutation screen in mouse.

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Bioentrepreneurship

Credits

3

Course Objectives

Research and business belong together and both are needed. In a rapidly developing life science industry, there is an urgent need for people who combine business knowledge with the understanding of science & technology. Bio-entrepreneurship, an interdisciplinary course, revolves around the central theme of how to manage and develop life science companies and projects. The objectives of this course are to teach students about concepts of entrepreneurship including identifying a winning business opportunity, gathering funding and launching a business, growing and nurturing the organization and harvesting the rewards.

Student Learning Outcomes

Students should be able to gain entrepreneurial skills, understand the various operations involved in venture creation, identify scope for entrepreneurship in biosciences and utilize the schemes promoted through knowledge centres and various agencies. The knowledge pertaining to management should also help students to be able to build up a strong network within the industry.

Unit I Innovation and entrepreneurship in bio-business 8 lectures

Introduction and scope in Bio-entrepreneurship. Types of bio-industries and competitive dynamics between the sub-industries of the bio-sector (e.g. pharmaceuticals vs. industrial biotech). Strategy and operations of bio-sector firms: Factors shaping opportunities for innovation and entrepreneurship in bio-sectors, and the business implications of those opportunities. Alternatives faced by emerging bio-firms and the relevant tools for strategic decision. Entrepreneurship development programs of public and private agencies (MSME, DBT, BIRAC, Make In India). strategic dimensions of patenting & commercialization strategies.

Unit II Bio markets - business strategy and marketing 8 lectures

Negotiating the road from lab to the market (strategies and processes of negotiation with financiers, government and regulatory authorities). Pricing strategy. Challenges in marketing in bio business (market conditions & segments; developing distribution channels, the nature, analysis and management of customer needs). Basic contract principles, different types of agreement and contract terms typically found in joint venture and development agreements, Dispute resolution skills.

Unit III Finance and accounting 8 lectures

Business plan preparation including statutory and legal requirements. Business feasibility study, financial management issues of procurement of capital and management of costs. Collaborations & partnership. Information technology.

Unit IV Technology management 8 lectures

Technology – assessment, development & upgradation. Managing technology transfer. Quality control & transfer of foreign technologies. Knowledge centres and Technology transfer agencies. Understanding of regulatory compliances and procedures (CDSCO, NBA, GCP, GLA, GMP).



Recommended Textbooks and References:

1. Adams, D.J., & Sparrow, J.C. (2008). *Enterprises for Life Scientists: Developing Innovation and Entrepreneurship in the Biosciences*. Bloxham: Scion.
2. Shinasaki, C. D. (2014). *Biotechnology Entrepreneurship: Starting, Managing, and Leading Biotech Companies*. Amsterdam: Elsevier. Academic Press is an imprint of Elsevier.

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1. Orrell, A., & Zucchella, A. *Business Modeling for Life Science and Biotech Companies: Creating Value and Competitive Advantage with the Milestone Bridge*. Routledge.
4. Jordan, J.F. (2014). *Innovation, Commercialization, and Start-Ups in Life Sciences*. London: CRC Press.
5. Desai, V. (2009). *The Dynamics of Entrepreneurial Development and Management*. New Delhi: Himalaya Pub. House.

Intellectual Property Rights, Biosafety and Bioethics

Credits

2

Course Objectives

The objectives of this course are:

- To provide basic knowledge on intellectual property rights and their implications in biological research and product development;
- To become familiar with India's IPR Policy;
- To learn biosafety and risk assessment of products derived from biotechnology and regulation of such products;
- To become familiar with ethical issues in biological research. This course will focus on consequences of biomedical research technologies such as cloning of whole organisms, genetic modifications, DNA testing.

Student Learning

Outcomes On completion

of this course, students should be able to:

- Understand the rationale for and against IPR and especially patents;
- Understand why India has adopted an IPR Policy and be familiar with broad outline of patent regulations;
- Understand different types of intellectual property rights in general and protection of products derived from biotechnology research and issues related to application and obtaining patents;
- Gain knowledge of biosafety and risk assessment of products derived from recombinant DNA research and environmental release of genetically modified organisms, national and international regulations;
- Understand ethical aspects related to biological, biomedical, health care and biotechnology research.

Unit I Introduction to IPR 5 lectures

Introduction to intellectual property; types of IP: patents, trademarks, copyright & related rights, industrial design, traditional knowledge, geographical indications, protection of new GMOs; International framework for the protection of IP; IP as a factor in R&D; IPs of relevance to biotechnology and few case studies; introduction to history of GATT, WTO, WIPO and TRIPS; plant variety protection and farmers rights act; concept of 'prior art'; invention in context of 'prior art'; patent databases - country-wise patent searches (USPTO, EPO, India); analysis and report formation.

Unit II Patenting 5 lectures

Basics of patents: types of patents; Indian Patent Act 1970; recent amendments; WIPO Treaties; Budapest Treaty, Patent Cooperation Treaty (PCT) and implications; procedure for filing a PCT application; role of a Country Patent Office; filing of a patent application; precautions before patenting-disclosure/non-disclosure - patent application-forms and guidelines including those of National Bio-diversity Authority (NBA) and other regulatory bodies, fee structure, time frames; types of patent applications: provisional and complete specifications; PCT and conventional patent applications; international patenting-requirement, procedures and costs; financial assistance for patenting-introduction to existing schemes; publication of patents-govt of India, status in Europe and US; patent infringement- meaning, scope, litigation, case studies and examples; commercialization of patented innovations; licensing - outright sale, licensing, royalty; patenting by research students and scientists-university/organizational rules in India and abroad, collaborative research - backward and forward IP; benefit/credit sharing among parties/community, commercial (financial) and non-commercial incentives.

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<p>Unit III Biosafety 5 lectures</p>	<p>Biosafety and Biosecurity - introduction; historical background; introduction to biological safety cabinets; primary containment for biohazards; biosafety levels; GRAS organisms; biosafety levels of specific microorganisms; recommended biosafety levels for infectious agents and infected animals; definition of GMOs & LMOs; principles of safety assessment of transgenic plants - sequential steps in risk assessment; concepts of familiarity and substantial equivalence; risk - environmental risk assessment and food and feed safety assessment; problem formulation - protection goals, compilation of relevant information, risk characterization and development of analysis plan; risk assessment of transgenic crops vs cisgenic plants or products derived from RNAi, genome editing tools.</p>
<p>Unit IV National and international regulations 5 lectures</p>	<p>International regulations - Cartagena protocol, OECD consensus documents and Codex Alimentarius; Indian regulations - EPA act and rules, guidance documents, regulatory framework - RCGM, GEAC, IBSC and other regulatory bodies; Draft bill of Biotechnology Regulatory authority of India - containments - biosafety levels and category of rDNA experiments; field trials - biosafety research trials - standard operating procedures - guidelines of state governments; GM labeling - Food Safety and Standards Authority of India (FSSAI).</p>
<p>Unit V Bioethics 5 lectures</p>	<p>Introduction, ethical conflicts in biological sciences - interference with nature, bioethics in health care - patient confidentiality, informed consent, euthanasia, artificial reproductive technologies, prenatal diagnosis, genetic screening, gene therapy, transplantation. Bioethics in research - cloning and stem cell research, Human and animal experimentation, animal rights/welfare, Agricultural biotechnology - Genetically engineered food, environmental risk, labeling and public opinion. Sharing benefits and protecting future generations - Protection of environment and biodiversity - biopiracy.</p>



Recommended Textbooks and References

1. Ganguli, P. (2001). *Intellectual Property Rights: Unleashing the Knowledge Economy*. New Delhi: Tata McGraw-Hill Pub.
2. *National IP Policy*, Department of Industrial Policy & Promotion, Ministry of Commerce, Govt.
3. *Complete Reference to Intellectual Property Rights Laws*. (2007). Snow White Publication Oct.
4. Kalne, H. (2010). *Bioethics: an Anthology*. Malden, MA: Blackwell.
5. Office of the Controller General of Patents, Design & Trademarks; Department of Industrial Policy & Promotion; Ministry of Commerce & Industry; Government of India. <http://www.ipindia.nic.in/>
6. Karen F. Greif and Jon F. Merz. *Current Controversies in the Biological Sciences - Case Studies of Policy Challenges from New Technologies*, MIT Press
7. World Trade Organisation. <http://www.wto.org>
8. World Intellectual Property Organisation. <http://www.wipo.int>
9. International Union for the Protection of New Varieties of Plants. <http://www.upov.int>
10. National Portal of India. <http://www.archive.india.gov.in>
11. National Biodiversity Authority. <http://www.nbaindia.org>
12. Recombinant DNA Safety Guidelines, 1990 Department of Biotechnology, Ministry of Science and Technology, Govt. of India. Retrieved from <http://www.envfor.nic.in/divisions/esurv/geac/annex-5.pdf>
13. Wolt, J. D., Keese, P., Raybould, A., Fitzpatrick, J. W., Burachik, M., Gray, A., Wu, F. (2009). Problem Formulation in the Environmental Risk Assessment for Genetically Modified Plants. *Transgenic Research*, 19(3), 425-436. doi:10.1007/s1248-009-9321-9
14. Craig, W., Tepfer, M., Degross, G., & Ripandelli, D. (2008). An Overview of General Features of Risk Assessments of Genetically Modified Crops. *Euphytica*, 164(3), 853-880. doi:10.1007/s10681-007-9643-8

Shukla

Rajsh

M



- 15. Guidelines for Safety Assessment of Foods Derived from Genetically Engineered Plants. 2008.
- 16. Guidelines and Standard Operating Procedures for Confined Field Trials of Regulated Genetically Engineered Plants. 2008. Retrieved from <http://www.ignrii.nic.in/guidelines1.asp>
- 17. Alonso, G. M. (2013). Safety Assessment of Food and Feed Derived from GM Crops: Using Problem Formulation to Ensure "Fit-for-Purpose" Risk Assessments. Retrieved from <http://biosafety.icgeb.org/inhousepublications/collection/biosafetyreviews>.

Project Proposal Preparation & Presentation

Credits

2

Course Objectives

The purpose of this course is to help students organize ideas, material and objectives for their dissertation and to begin development of communication skills and to prepare the students to present their topic of research and explain its importance to their fellow classmates and teachers.

Student Learning Outcomes

Students should be able to demonstrate the following abilities:

- Formulate a scientific question;
- Present scientific approach to solve the problem;
- Interpret, discuss and communicate scientific results in written form;
- Gain experience in writing a scientific proposal;
- Learn how to present and explain their research findings to the audience effectively.

Syllabus

Project Proposal Preparation

Selection of research lab and research topic: Students should first select a lab wherein they would like to pursue their dissertation. The supervisor or senior researchers should be able to help the students to read papers in the areas of interest of the lab and help them select a topic for their project. The topic of the research should be hypothesis driven.

Review of literature: Students should engage in systematic and critical review of appropriate and relevant information sources and appropriately apply qualitative and/or quantitative evaluation processes to original data, keeping in mind ethical standards of conduct in the collection and evaluation of data and other resources.

Writing Research Proposal: With the help of the senior researchers, students should be able to discuss the research questions, goals, approach, methodology, data collection, etc. Students should be able to construct a logical outline for the project including analysis steps and expected outcomes and prepare a complete proposal in scientific proposal format for dissertation.

Syllabus

Poster Presentation

Students will have to present the topic of their project proposal after few months of their selection of the topic. They should be able to explain the novelty and importance of their research topic.

Syllabus

Oral Presentation

At the end of their project, presentation will have to be given by the students to explain work done by them in detail. Along with summarizing their findings they should also be able to discuss the future expected outcome of their work.

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Laboratory VI: Bioprocess Engineering & Technology

Credits



Course Objectives

The objectives of this laboratory course are to provide hands-on training to students in upstream and downstream unit operations.

Student Learning Outcomes

Students should be able to:

- Investigate, design and conduct experiments, analyze and interpret data, and apply the laboratory skills to solve complex bioprocess engineering problems;
- Apply skills and knowledge gained will be useful in solving problems typical of bio industries and research.

Syllabus

- Basic Microbiology techniques
 - Scale up from frozen vial to agar plate to shake flask culture.
 - Instrumentation: Microplate reader, spectrophotometer, microscopy.
 - Isolation of microorganisms from soil samples.
- Experimental set-up
 - Assembly of bioreactor and sterilization.
 - Growth kinetics.
 - Substrate and product inhibitions.
 - Measurement of residual substrates.
- Data Analysis
 - Introduction to Metabolic Flux Analysis (MFA).
- Fermentation
 - Batch.
 - Fed-batch.
 - Continuous.
- Unit operations
 - Microfiltrations: Separation of cells from broth.
 - Bioseparations: Various chromatographic techniques and extractions.
- Biomanalytics
 - Analytical techniques like HPLC, FPLC, GC, GC-MS etc. for measurement of amounts of products/substrates.



Recommended Textbooks and References:

- Stuler, M. L., & Kargi, F. (2002). *Bioprocess Engineering: Basic Concepts*. Upper Saddle River, NJ: Prentice Hall.
- Stanbury, P. F., & Whitaker, A. (2010). *Principles of Fermentation Technology*. Oxford: Pergamon Press.
- Blanch, H. W., & Clark, D. S. (1997). *Biochemical Engineering*. New York: M. Dekker.
- Bailey, J. E., & Ollis, D. F. (1986). *Biochemical Engineering Fundamentals*. New York: McGraw-Hill.
- El-Mansi, M., & Bryoz, C. F. (2007). *Fermentation Microbiology and Biotechnology*. Boca Raton: CRC/Taylor & Francis.

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Laboratory VII: Bioinformatics

Credits



Course Objectives

The aim of this course is to provide practical training in bioinformatic methods including accessing major public sequence databases, use of different computational tools to find sequences, analysis of protein and nucleic acid sequences by various software packages.

Student Learning Outcomes

On completion of this course, students should be able to:

- 1. Describe contents and properties of most important bioinformatics databases;
- 2. Perform text- and sequence-based searches and analyze and discuss results in light of molecular biological knowledge;
- 3. Explain major steps in pairwise and multiple sequence alignment, explain principle and execute pairwise sequence alignment by dynamic programming;
- 4. Predict secondary and tertiary structures of protein sequences.

Syllabus

1. Using NCBI and Uniprot web resources.
2. Introduction and use of various genome databases.
3. Sequence information resource: Using NCBI, EMBL, Genbank, Enzet, Swissprot, TrEMBL, UniProt.
4. Similarity searches using tools like BLAST and interpretation of results.
5. Multiple sequence alignment using ClustalW.
6. Phylogenetic analysis of protein and nucleotide sequences.
7. Use of gene prediction methods (GRAIL, Genscan, Glimmer).
8. Using RNA structure prediction tools.
9. Use of various primer designing and restriction site prediction tools.
10. Use of different protein structure prediction databases (PDB, SCOP, CATH).
11. Construction and study of protein structures using Deepview/PyMol.
12. Homology modelling of proteins.
13. Use of tools for mutation and analysis of the energy minimization of protein structures.
14. Use of miRNA prediction, designing and target prediction tools.

Semester Four

Dissertation

Credits

24

(Semester III: 4 Credits
Semester IV: 20 Credits)

Course Objectives

The objectives of this course are to prepare the students to adapt to the research environment and understand how projects are executed in a research laboratory. It will also enable students to learn practical aspects of research and train students in the art of analysis and thesis writing.

Student Learning Outcomes

Students should be able to learn how to select and defend a topic of their research, how to effectively plan, execute, evaluate and discuss their experiments. Students should be able to demonstrate considerable improvement in the following areas:

- 1. In-depth knowledge of the chosen area of research;
- 2. Capability to critically and systematically integrate knowledge to identify issues that must be addressed within framework of specific thesis.
- 3. Competence in research design.

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- Capability to create, analyse and critically evaluate different technical solutions.
 - Ability to conduct research independently.
 - Ability to perform analytical techniques/experimental methods.
 - Project management skills.
 - Report writing skills.
 - Problem solving skills.
 - Communication and interpersonal skills.

Syllabus

Planning & performing experiments

Based on the project proposal submitted in earlier semester, students should be able to plan, and engage in, an independent and sustained critical investigation and evaluate a chosen research topic relevant to biological sciences and society. They should be able to systematically identify relevant theory and concepts, relate these to appropriate methodologies and evidence, apply appropriate techniques and draw appropriate conclusions. Senior researchers should be able to train the students such that they can work independently and are able to understand the aim of each experiment performed by them. They should also be able to understand the possible outcomes of each experiment.

Syllabus

Thesis writing

At the end of their project, thesis has to be written giving all the details such as aim, methodology, results, discussion and future work related to their project. Students may aim to get their research findings published in a peer-reviewed journal. If the research findings have application-oriented outcomes, the students may file patent application.

Recommended Electives

Biological Imaging

Credits

2

Course Objectives

The objectives of this course are to provide complete overview of state-of-art live-cell imaging techniques using microscopes currently available in literature. Live-cell imaging techniques allow real-time examination of almost every aspect of cellular function under normal and experimental conditions. With live-cell imaging experiments, main challenges are to keep cells alive and healthy over a period of time. The growing number of live-cell imaging techniques means one can obtain greater amounts of information without stressing out cells.

Student Learning Outcomes

On completion of this course, students shall be able to gain complete overview of super-resolution field from fundamentals to state-of-art methods and applications in biomedical research. The students shall learn the comparative advantages and disadvantages of each technique, covers all key techniques in field of biomedical science. The students shall also learn how to use new tools to increase resolution in sub-nanometer-scale images of living cells and tissue, which leads to new information about molecules, pathways and dynamics and state-of-the-art examples of applications using microscopes.

Unit 1

Widefield fluorescent microscopy
3 lectures

One of the most basic techniques for live-cell imaging is widefield fluorescent microscopy. Standard inverted research grade microscopes can yield valuable results if you are imaging the entire cell, large regions of interest, such as organelles, or very thin tissue sections (less than 5 micrometer). In widefield, a CCD camera is usually used to

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capture images and the epi-fluorescence illumination source can be a mercury lamp, xenon lamp, LED's, etc. Each of light sources require carefully matched interference filters for specific excitation and emission wavelengths of your fluorophore of interest. With widefield microscopy, your specimen is only exposed to excitation light for relatively short time periods as the full aperture of emission light is collected by the objectives. Widefield fluorescence microscopy can be used in combination with other common contrast techniques such as phase contrast and differential interference contrast (DIC) microscopy. This combination is useful when performing live-cell imaging to examine general cell morphology or viability while also imaging regions of interest within cells.

Unit II
Confocal laser scanning microscopy (CLSM)
3 lectures

CLSM has ability to eliminate out-of-focus light and information. It is also possible to obtain optical aerial sections from thicker specimens. A conjugate pinhole in optical path of confocal microscope prevents fluorescence from outside of focal plane from being collected by photomultiplier detector or imaged by camera. In CLSM, a single pinhole (and single focused laser spot) is scanned across specimen by scanning system. This spot forms a reflected epi-fluorescence image back on original pinhole. When specimen is in focus, fluorescent light from it passes through pinhole to detector. Any out-of-focus light is defocused at pinhole and very little of this signal passes through to detector meaning that background fluorescence is greatly reduced. The pinhole acts as a spatial filter for emission light from the specimen.

Unit III
Spinning disc confocal microscopy (SDCM)
2 lectures

This method utilizes a 'Nipkow Disc' which is a mechanical coprocessor which has a series of thousands of tiny holes or pinholes arranged in a spiral pattern. Each illuminated pinhole on disc is imaged by microscope objective to a diffraction-limited spot on region of interest on specimen. The emission from fluorophores passes back through Nipkow disc pinholes and can be observed and captured by a CCD camera. The effect of spinning disc is that many thousands of points on specimen are simultaneously illuminated. Using SDCM to examine a specimen means that real-time imaging (30-frames-per-second or faster) can be achieved, which is extremely useful if you are looking at dynamic changes within living cells over a wide spectrum of time-scales.

Unit IV
Light-sheet fluorescence microscopy (LSFM, or SPIM)
2 lectures

This method enables one to perform live-cell imaging of whole embryos, tissues and cell spheroids *in vivo* in a gentle manner with high temporal resolution and in three dimensions. One is able to track cell movement over extended periods of time and follow development of organs and tissues on a cellular level. The next evolution of light-sheet fluorescence microscopy, termed lattice light-sheet microscopy as developed by Eric Betzig (Nobel Prize Laureate 2014 for PALM super-resolution microscopy) will even allow live-cell imaging with super-resolved *in vivo* cellular localization capabilities.

Unit V
Super-resolved fluorescence microscopy
8 lectures

Super-Resolution in a Standard Microscope: From Fast Fluorescence Imaging to Molecular Diffusion Laws in Live Cells; Photoswitching Fluorophores in Super-Resolution Fluorescence Microscopy; Image Analysis for Single-Molecule Localization Microscopy; Deconvolution of Nanoscopic Images; Super-Resolution Fluorescence Microscopy of the Nanoscale Organization in cells; Correlative Live-Cell and Super-Resolution Microscopy and Its Biological Applications; SAX Microscopy and Its Application to Imaging of 3D-Cultured Cells; Quantitative Super-Resolution Microscopy for Cancer Biology and Medicine.

Unit VI
Re-scan confocal microscopy
4 lectures

Structured Illumination Microscopy; Correlative Nanoscopy; AFM Super-Resolution (STED-STORM); Stochastic Optical Fluctuation Imaging.

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Recommended Textbooks and References:

1. Rajagopal Vadivambal, Digvir S. Jayas, (2015). *Bio-Imaging: Principles, Techniques, and Applications*. ISBN 9781466593671 -CAT# K20618.
2. Alberto Diaspro, Marc A. M. J. van Zandvoort, (2016). *Super-Resolution Imaging in Biomedicine*. ISBN 9781482244342 -CAT# K23483.
3. Tsjies, Douglas, Roth, Jürgen (Eds.) (2012). *Cell Imaging Techniques Methods and Protocols*. ISBN 978-1-62703-056-4.

Computational Biology

Credits

2

Course Objectives

The objective of this course is to provide students with theory and practical experience of essentials to aid for genomic, proteomic and metabolomics courses and drug design program.

Student Learning Outcomes

On completion of this course, the students are expected to:

- 1. Develop an understanding of the basic theory of these computational tools;
- 2. Develop required database extraction, integration, coding for computational tools and methods necessary for all Omics;
- 3. Create hypothesis for investigating specific contemporary biological questions, provide help to experiment with or develop appropriate tools;
- 4. Critically analyze and interpret results of their study with respect to whole systems.

Unit I

Introduction to computational biology basics and biological databases

4 lectures

Computers in biology and medicine; Overview of biological databases, nucleic acid & protein databases, primary, secondary, functional, composite, structural classification database, Sequence formats & storage, Access databases, Extract and create sub databases, limitations of existing databases.

Unit II

Pairwise and multiple sequence alignments

6 lectures

Local alignment, Global alignment, Scoring matrices - PAM, BLOSUM, Gaps and penalties, Dot plots. Dynamic programming approach: Needleman and Wunsch Algorithm, Smith and Waterman Algorithm, Hidden Markov Model: Viterbi Algorithm. Heuristic approach: BLAST, FASTA. Building Profiles, Profile based functional identification.

Unit III

Genome analysis

6 lectures

Polymorphisms in DNA sequence, Introduction to Next Generation Sequencing technologies, Whole Genome Assembly and challenges, Sequencing and analysis of large genomes, Gene prediction, Functional annotation, Comparative genomics, Probabilistic functional gene networks, Human genome project, Genomics and crop improvement. Study available GWAS, ENCODE, HUGO projects, extract and build sub databases; Visualization tools including Artemis and Vista for genome comparison; Functional genomics case studies.

Unit IV

Structure visualization

3 lectures

Retrieving and drawing structures, Macromolecule viewing platforms, Structure validation and correction, Structure optimization, Analysis of ligand-protein interactions; Tools such as PyMol or VMD.

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Unit V
Molecular modelling
8 lectures

Significance and need, force field methods, energy, buried and exposed residues; side chains and neighbours; fixed regions; hydrogen bonds; mapping properties onto surfaces; RMS fit of conformers and protein chains, assigning secondary structures; sequence alignment: methods, evaluation, scoring; protein curation: backbone construction and side chain addition; different types of protein chain modelling: *ab initio*, homology, hybrid, loop; Template recognition and alignments; Modelling parameters and considerations; Model analysis and validation; Model optimization; Substructure manipulations, annealing, protein folding and model generation; loop generating methods; loop analysis; Analysis of active sites using different methods in studying protein-protein interactions.

Unit VI
Structure-based drug development
8 lectures

Molecular docking: Types and principles, Semi-flexible docking, Flexible docking; Ligand and protein preparation, Macromolecule and ligand optimization, Ligand conformations, Clustering; Analysis of docking results and validation with known information. Extra-precision docking platforms, Use of Small-molecule libraries, Natural compound libraries for virtual high throughput screenings.

Unit VII
Ligand-based drug development
8 lectures

Quantitative structure activity relationships; Introduction to chemical descriptors like 2D, 3D and Group-based, Radar plots and contribution plots and Activity predictions, Pharmacophore modeling, Pharmacophore-based screenings of compound library, analysis and experimental validation.



Recommended Textbooks and References:

1. Mount, D.W. (2001). *Bioinformatics: Sequence and Genome Analysis*. Cold Spring Harbor, NY: Cold Spring Harbor Laboratory Press.
2. Bourne, P.E., & Gu, J. (2009). *Structural Bioinformatics*. Hoboken, NJ: Wiley-Liss.
3. Lesk, A.M. (2004). *Introduction to Protein Science: Architecture, Function, and Genomics*. Oxford: Oxford University Press.
4. Campbell, M. & Heyer, L.J. (2006). *Discovering Genomics, Proteomics and Bioinformatics*. Pearson Education.
5. Oprea, T. (2005). *Cheminformatics in Drug Discovery*, Volume 23, Wiley Online Library.
6. Gasteiger, J. & Engel, T. (2003). *Cheminformatics: a Textbook*, Wiley Online Library.

Drug Discovery and Development

Credits

2

Course Objectives

This course will give a broad overview of research and development carried out in industrial setup towards drug discovery.

Student Learning Outcomes

On completion of this course, students should be able to understand basics of R&D in drug discovery and should be able to apply knowledge gained in respective fields of pharmaceutical industry.

Unit I
Target identification and molecular modelling
7 lectures

Identification of target or drug leads associated with a particular disease by a number of different techniques including combinations of molecular modeling, combinatorial libraries and high-throughput screening (HTS); Conceptualizing the automation of the HTS process and the importance of bioinformatics and data processing in identification of lead compounds; Rational drug design, based on understanding the three-dimensional

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structures and physicochemical properties of drugs and receptors; Modelling drug/receptor interactions with the emphasis on molecular mechanisms, molecular dynamics simulations and homology modelling; Conformational sampling, macromolecular folding, structural bioinformatics, receptor-based and ligand-based design and docking methods, in silico screening of libraries, semi-empirical and ab-initio methods, QSAR methods, molecular diversity, design of combinatorial libraries of drug-like molecules, macromolecular and chemical databases.

Unit II Lead optimization 5 lectures	Identification of relevant groups on a molecule that interact with a receptor and are responsible for biological activity; Understanding structure activity relationship; Structure modification to increase potency and therapeutic index; Concept of quantitative drug design using Quantitative structure-activity relationship models (QSAR models) based on the fact that the biological properties of a compound are a function of its physicochemical parameters such as solubility, lipophilicity, electronic effects, ionization, stereochemistry, etc.; Bioanalytical assay development in support of in vitro and in vivo studies (LC/MS/MS, GC/MS and ELISA).
Unit III Preclinical development 5 lectures	Principles of drug absorption, drug metabolism and distribution - intestinal absorption, metabolic stability, drug-drug interactions, plasma protein binding assays, metabolic profile studies, Principles of toxicology, Experimental design for preclinical and clinical PK/PD/TK studies, Selection of animal model; Regulatory guidelines for preclinical PK/PD/TK studies; Scope of GLP, SOP for conduct of clinical & non clinical testing, control on animal house, report preparation and documentation Integration of non-clinical and preclinical data to aid design of clinical studies.
Unit IV Drug manufacturing 4 lectures	Requirements of GMP implementation, Documentation of GMP practices, CoA, Regulatory certification of GMP, Quality control and Quality assurance, concept and philosophy of TQM, ICH and ISO 9000; ICH guidelines for Manufacturing, Understanding Impurity Qualification Data, Stability Studies.
Unit V Clinical trial design 4 lectures	Objectives of Phase I, II, III and IV clinical studies, Clinical study design, enrollment, sites and documentation, Clinical safety studies: Adverse events and adverse drug reactions, Clinical PK, pharmacology, drug-drug interaction studies, Statistical analysis and documentation.
Unit VI Fundamentals of regulatory affairs and bioethics 4 lectures	Global Regulatory Affairs and different steps involved, Regulatory Objectives, Regulatory Agencies; FDA guidelines on IND and NDA submissions, Studies required for IND and NDA submissions for oncology, HIV, cardiovascular indications, On-label vs. off-label drug use GCP and Requirements of GCP Compliance, Ethical issues and Compliance to current ethical guidelines, Ethical Committees and their set up, Animal Ethical issues and compliance.



Recommended Textbooks and References:

1. Krosgaard-Larsen et al. *Textbook/Drug Design and Discovery*, 4th Edition. CRC Press.
2. Kuhse, H. (2010), *Bioethics: an Anthology*. Malden, MA: Blackwell.
3. Nally, J. D. (2006) *GMP for Pharmaceuticals*, 6th edition. CRC Press
4. Brody, T. (2016) *Clinical Trials: Study Design, Endpoints and Biomarkers*, Drug Safety, and FDA and ICH Guidelines. Academic Press.

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Environmental Biotechnology

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Course Objectives

This course aims to introduce fundamentals of Environmental Biotechnology. The course will introduce major groups of microorganisms- tools in biotechnology and their most important environmental applications. The environmental applications of biotechnology will be presented in detail and will be supported by examples from the national and international literature.

Student Learning Outcomes

On completion of course, students will be able to understand use of basic microbiological, molecular and analytical methods, which are extensively used in environmental biotechnology.

Unit I Introduction to environment 8 lectures

Introduction to environment; pollution and its control; pollution indicators; waste management: domestic, industrial, solid and hazardous wastes; strain improvement; Biodiversity and its conservation; Role of microorganisms in geochemical cycles; microbial energy metabolism, microbial growth kinetics and elementary chemostat theory, relevant microbiological processes, microbial ecology.

Unit II Bioremediation 6 lectures

Bioremediation: Fundamentals, methods and strategies of application (biostimulation, bioaugmentation) – examples, bioremediation of metals (Cr, As, Se, Hg), radionuclides (U, Te), organic pollutants (PAHs, PCBs, Pesticides, TNT etc.), technological aspects of bioremediation (in situ, ex situ).

Unit III Role of microorganisms in bioremediation 6 lectures

Application of bacteria and fungi in bioremediation: White rot fungi vs specialized degrading bacteria: examples, uses and advantages vs disadvantages; Phytoremediation: Fundamentals and description of major methods of application (phytoaccumulation, phytovolatilization, rhizofiltration phytostabilization).

Unit IV Biotechnology and agriculture 11 lectures

Bioinsecticides: *Bacillus thuringiensis*, Baculoviruses, uses, genetic modifications and aspects of safety in their use; Biofungicides: Description of mode of actions and mechanisms (e.g. *Trichoderma*, *Pseudomonas fluorescens*); Biofertilizers: Symbiotic systems between plants – microorganisms (nitrogen fixing symbiosis, mycorrhiza fungi symbiosis), Plant growth promoting rhizobacteria (PGPR) – uses, practical aspects and problems in application.

Unit V Biofuels 11 lectures

Environmental Biotechnology and biofuels: biogas; bioethanol; biodiesel; biohydrogen; Description of the industrial processes involved, microorganisms and biotechnological interventions for optimization of production; Microbiologically enhanced oil recovery (MEOR); Biobleaching of metals; Production of bioplastics; Production of biosurfactants; bioemulsifiers; Paper production: use of xylanases and white rot fungi.



Recommended Textbooks and References:

1. G. M. Evans and J. C. Furlong (2003), *Environmental Biotechnology: Theory and Applications*, Wiley Publishers.
2. B. Ritman and P. L. McCarty (2000), *Environmental Biotechnology: Principle & Applications*, 2nd Ed., McGraw Hill Science.
3. Scragg A., (2005) *Environmental Biotechnology*, Pearson Education Limited.
4. J. S. Devlany, M. A. Deshusses and T. S. Webster, (1998), *Biofiltration for Air Pollution Control*, CRC Press.
5. H. J. Rehm and G. Reed, (2001), *Biotechnology - A Multi-volume Comprehensive Practice*, Vol. 11, 2nd Ed., VCH Publishers Inc.

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6 H. S. Peavy, D. R. Rowe and G. Tchobanoglous, (2013), *Environmental Engineering*, McGraw-Hill Inc.

Microbial Technology

Credits

2

Course Objectives

The objectives of this course are to introduce students to developments/ advances made in field of microbial technology for use in human welfare and solving problems of the society.

Student Learning Outcomes

On completion of this course, students would develop deeper understanding of the microbial technology and its applications.

Unit I

Introduction to microbial technology 8 lectures

Microbial technology in human welfare; Isolation and screening of microbes important for industry – advances in methodology and its application; Advanced genome and epigenome editing tools (e.g., engineered zinc finger proteins, TALEs/TALENs, and the CRISPR/Cas9 system as nucleases for genome editing, transcription factors for epigenome editing, and other emerging tools) for manipulation of useful microbes/ strains and their applications; Strain improvement to increase yield of selected molecules, e.g., antibiotics, enzymes, biofuels.

Unit II

Environmental applications of microbial technology 6 lectures

Environmental application of microbes; Ore leaching; Biodegradation - biomass recycle and removal; Bioremediation - toxic waste removal and soil remediation; Global Biogeochemical cycles; Environment sensing (sensor organisms/ biological sensors); International and National guidelines regarding use of genetically modified organisms in environment, food and pharmaceuticals.

Unit III

Pharmaceutical applications of microbial technology 8 lectures

Recombinant protein and pharmaceuticals production in microbes – common bottlenecks and issues (technical/operational, commercial and ethical); Attributes required in industrial microbes (*Streptomyces* sp., Yeast) to be used as efficient cloning and expression hosts (biologicals production); Generating diversity and introduction of desirable properties in industrially important microbes (*Streptomyces*/Yeast); Microbial cell factories; Downstream processing approaches used in industrial production process (*Streptomyces* sp., Yeast).

Unit IV

Food applications of microbial technology 7 lectures

Application of microbes and microbial processes in food and healthcare industries - food processing and food preservation, antibiotics and enzymes production, microbes in targeted delivery application – drugs and vaccines (bacterial and viral vectors); Non-recombinant ways of introducing desirable properties in Generally recognized as safe (GRAS) microbes to be used in food (e.g., Yeast) - exploiting the existing natural diversity or the artificially introduced diversity through conventional acceptable techniques (mitogenesis, protoplast fusion, breeding, genome shuffling, directed evolution etc.).

Unit V

Advances in microbial technology 8 lectures

Microbial genomics for discovery of novel enzymes, drugs/ antibiotics; Limits of microbial genomics with respect to use in human welfare; Metagenomics and metatranscriptomics – their potential, methods to study and applications/use (animal and plant health, environmental clean-up, global nutrient cycles & global sustainability, understanding evolution); Global metagenomics initiative - surveys/projects and outcome, metagenomic library construction and functional screening in suitable hosts – tools and techniques for discovery/identification of novel enzymes, drugs (e.g., protease, antibiotic) etc.

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Recommended Textbooks and References:

1. Lee, Y.K. (2013). *Microbial Biotechnology: Principles and Applications*. Hackensack, NJ: World Scientific.
2. Moo-Young, M. (2011). *Comprehensive Biotechnology*. Amsterdam: Elsevier.
3. Nelson, K. E. (2015). *Encyclopedia of Metagenomics: Genes, Genomes and Metagenomes: Basics, Methods, Databases and Tools*. Boston, MA: Springer US.
4. *The New Science of Metagenomics: Revealing the Secrets of Our Microbial Planet*. (2007). Washington, D.C.: National Academies Press.
5. Journals: (a) Nature, (b) Nature Biotechnology, (c) Applied microbiology and biotechnology, (d) Trends in Biotechnology, (e) Trends in Microbiology, (f) Current opinion in Microbiology, (g) Biotechnology Advances, (h) Genome Research
6. Websites: <http://jgi.doe.gov/our-science/>

Protein Engineering

Credits

2

Course Objectives

The aim of this course is to introduce methods and strategies commonly used in protein engineering.

Student Learning Outcomes

On completion of this course, students should be able to:

- 1. Analyse structure and construction of proteins by computer-based methods;
- 2. Describe structure and classification of proteins;
- 3. Analyse purity and stability of proteins and explain how to store them in best way;
- 4. Explain how proteins can be used for different industrial and academic purposes such as structure determination, organic synthesis and drug design.

Unit I

Introduction to protein engineering
5 lectures

Protein engineering – definition, applications; Features or characteristics of proteins that can be engineered (definition and methods of study) – affinity and specificity; Spectroscopic properties; Stability to changes in parameters as pH, temperature and amino acid sequence, aggregation propensities, etc. Protein engineering with unnatural amino acids and its applications.

Unit II

Stability of protein structure
5 lectures

Methods of measuring stability of a protein; Spectroscopic methods to study physicochemical properties of proteins: far-UV and near-UV CD; Fluorescence; UV absorbance; ORD; Hydrodynamic properties – viscosity, hydrogen-deuterium exchange; Brief introduction to NMR spectroscopy – emphasis on parameters that can be measured/obtained from NMR and their interpretation.

Unit III

Applications
5 lectures

Forces stabilizing proteins – Van der Waals, electrostatic, hydrogen bonding and weakly polar interactions, hydrophobic effects; Entropy – enthalpy compensation; Experimental methods of protein engineering: directed evolution like gene site saturation mutagenesis; Modular shuffling; Guided protein recombination, etc., Optimization and high throughput screening methodologies like GigaMatrix, High throughput microplate screens etc., Application to devices with bacteriorhodopsin as an example; Engineering antibody affinity by yeast surface display; Applications to vaccines, Peptidomimetics and its use in drug discovery.

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Unit IV
Computational approaches
5 lectures

Computational approaches to protein engineering: sequence and 3D structure analysis, Data mining, Ramachandran map, Mechanism of stabilization of proteins from psychrophiles and thermophiles vs those from mesophiles; Proteindesign, Directed evolution for protein engineering and its potential.

Unit V
Case studies
1 lecture

Case Studies.



Recommended Textbooks and References:

1. Edited by T. Creighton, (1997), *Protein Structure: A Practical Approach*, 2nd Edition, Oxford university press.
2. Cleland and Craik, (2006), *Protein Engineering, Principles and Practice*, Vol 7, Springer Netherlands.
3. Mueller and Arndt, *Protein Engineering Protocols*, 1st Edition, Humana Press.
4. Ed. Robertson DE, Noel JP, (2004), *Protein Engineering, Methods in Enzymology*, 388, Elsevier Academic Press.
5. J Kyte, (2006), *Structure in Protein Chemistry*, 2nd Edition, Garland publishers.

Nano-biotechnology

Credits

2

Course Objectives

The course aims at providing a general and broad introduction to multi-disciplinary field of nanotechnology. It will familiarize students with the combination of the top-down approach of microelectronics and micromechanics with the bottom-up approach of chemistry/biochemistry, a development that is creating new and exciting cross-disciplinary research fields and technologies. The course will also give an insight into complete systems where nanotechnology can be used to improve our everyday life.

Student Learning Outcomes

On successful completion of this course, students should be able to describe basic science behind the properties of materials at nanometre scale, and the principles behind advanced experimental and computational techniques for studying nanomaterials.

Unit I
Introduction to nanobiotechnology
5 lectures

Introduction to Nanobiotechnology; Concepts, historical perspective; Different formats of nanomaterials and applications with example for specific cases; Cellular Nanostructures; Nanopores; Biomolecular motors; Bio-inspired Nanostructures, Synthesis and characterization of different nanomaterials.

Unit II
Nano – films
5 lectures

Thin films; Colloidal nanostructures; Self Assembly; Nanovesicles; Nanospheres; Nanocapsules and their characterisation.

Unit III
Nano – particles
5 lectures

Nanoparticles for drug delivery, concepts, optimization of nanoparticle properties for suitability of administration through various routes of delivery, advantages, strategies for cellular internalization and long circulation, strategies for enhanced permeation through various anatomical barriers.

Unit IV
Applications of nano-particles
5 lectures

Nanoparticles for diagnostics and imaging (theranostics); concepts of smart stimuli responsive nanoparticles, implications in cancer therapy, nanodevices for biosensor development.

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Unit V Nano-materials 5 lectures	Nanomaterials for catalysis, development and characterization of nanobiocatalysts, application of nanoscaffolds in synthesis, applications of nanobiocatalysis in the production of drugs and drug intermediates.
Unit VI Nano-toxicity 5 lectures	Introduction to Safety of nanomaterials, Basics of nanotoxicity, Models and assays for Nanotoxicity assessment; Fate of nanomaterials in different strata of environment; Ecotoxicity models and assays; Life Cycle Assessment, containment.



Recommended Textbooks and References:

1. Gero Decher, Joseph B. Schlenker, (2003); *Multilayer Thin Films: Sequential Assembly of Nanocomposite Materials*, Wiley-VCH Verlag GmbH & Co. KGaA
2. David S. Goetsell, (2004); *Bionanotechnology: Lessons from Nature*, Wiley-Liss
3. Neelina H. Malach (2005); *Biomedical Nanotechnology*, CRC Press
4. Greg T. Hermanson, (2013); *Bioconjugate Techniques*, (3rd Edition); Elsevier
5. Recent review papers in the area of Nanomedicine.

Vaccines

Credits

2

Course Objectives

This course will provide students with an overview of current developments in different areas of vaccines.

Student Learning Outcomes

By the end of this course, students should be able to:

- 1. Understand fundamental concepts of human immune system and basic Immunology;
- 2. Differentiate and understand immune responses in relation to infection and vaccination;
- 3. Understand requirement and designing of different types of vaccines;
- 4. Understand importance of conventional and new emerging vaccine technologies.

Unit I Fundamentals of immune system 6 lectures	Overview of Immune system; Human Immune system; Effectors of immune system; Innate & Adaptive Immunity; Activation of the Innate Immunity; Adaptive Immunity; T and B cells in adaptive immunity; Immune response in infection; Correlates of protection.
Unit II Immune response to infection 9 lectures	Protective immune response in bacterial; viral and parasitic infections; Primary and Secondary immune responses during infection; Antigen presentation and Role of Antigen presenting cells; Dendritic cells in immune response; Innate immune response; Humoral (antibody mediated) responses; Cell mediated responses: role of CD4+ and CD8+ T cells; Memory responses; Memory and effector T and B cells, Generation and Maintenance of memory T and B cells.
Unit III Immune response to vaccination 8 lectures	Vaccination and immune response; Adjuvants in Vaccination; Modulation of immune responses: Induction of Th1 and Th2 responses by using appropriate adjuvants and antigen delivery systems - Microbial adjuvants, Liposomal and Microparticles as delivery systems; Chemokines and cytokines; Role of soluble mediators in vaccination; Oral immunization and Mucosal Immunity.
Unit IV Vaccine types & design 3 lectures	History of vaccines, Conventional vaccines; Bacterial vaccines; Viral Vaccines; Vaccines based on routes of administration: parenteral, oral, mucosal; Live attenuated and inactivated vaccine; Subunit Vaccines and Toxoids; Peptide Vaccine.

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Unit V
Vaccine technologies
4 lectures

New Vaccine Technologies; Rationally designed Vaccines; DNA Vaccination; Mucosal vaccination; New approaches for vaccine delivery; Engineering virus vectors for vaccination; Vaccines for targeted delivery (Vaccine Delivery systems); Disease specific vaccine design; Tuberculosis Vaccine; Malaria Vaccine; HIV/AIDS vaccine; New emerging diseases and vaccine needs (Ebola, Zika).



Recommended Textbooks and References:

1. Janeway, C. A., Traves, P., Walport, M., & Shlomchik, M. J. (2005). *Immunobiology: The Immune System in Health and Disease*. USA: Garland Science Pub.
2. Kindt, T. J., Osborne, B. A., Goldby, R. A., & Kuby, J. (2013). *Ruby Immunology*. New York: W. H. Freeman.
3. Kaufmann, S. H. (2004). *New Vaccination Strategies*. Weinheim: Wiley-VCH.
4. Journal Articles (relevant issues) from: Annual Review of Immunology, Annual Review of Microbiology, Current Opinions in Immunology, Nature Immunology, Expert review of vaccines.

Bhhatt
9/7/2020

Roy

9/7/20



Medical Microbiology and Infection Biology

Credits

3

Course Objectives

This course will provide a perspective and exposure to medical aspects of bacteriology, virology, mycology, parasitology and infectious diseases along with concepts of symptoms, pathogenesis, transmission, prophylaxis and control, a conceptual understanding of host – pathogen interactions using well characterized systems as examples. The student should have a good grasp of disease causing microbes and their interactions with host.

Student Learning Outcomes

On completion of this course, students should be able to:

- Compare and contrast different microbial diseases, including properties of different types of pathogens, and mechanisms of pathogenesis;
- Summarize role of host in infectious disease, including natural barriers to infection, innate and acquired immune responses to infection, and inflammation;
- Compare and contrast experimental approaches for identifying virulence genes and advantages/disadvantages of each approach for specific pathogens.

Unit I

Bacterial diseases

8 lectures

Normal microflora (microbiome) of human body and its role – Skin, mouth and respiratory tract, intestinal tract, urogenital tract; Pathogenesis and virulence factors – Koch's postulates, Adherence and invasion, Toxins, Enzymes, Antiphagocytic factors, Antigenic heterogeneity, Iron acquisition; *Bacillus anthracis*, *Clostridium* spp., *Corynebacterium diphtheriae*; *E. coli*, *Vibrio cholerae*, *Helicobacter pylori*, *Salmonella typhi* and *paratyphi*, *Shigella dysenteriae*; *Listeria monocytogenes*, *Mycobacterium* spp., Rickettsial diseases; *Haemophilus influenzae*, *Bordetella pertussis*, Brucellosis, Streptococcal and Staphylococcal infections; Antibacterial chemotherapy (with examples of antibiotics) - Inhibition of cell wall synthesis, inhibition of cell membrane function, inhibition of protein and nucleic acid synthesis, antimetabolites; Drug resistance - origin (genetic and non-genetic), mechanisms, antimicrobial activity in vitro and in vivo, Multi-drug resistance and its mechanisms e.g. MDR-TB.

Unit II

Viral diseases

7 lectures

Viral Pathogenesis - Routes of entry, Viral spread (local and systemic infection), Viral persistence (chronic and latent infection); Polio, Chicken pox, Mumps, Measles, Rubella; Viral hemorrhagic fever, viral encephalitis, Dengue and Yellow fever; Influenza virus infection (emphasis on Avian and swine flu), Rabies and Prion diseases; Hepatitis and Human Cancer viruses; Emerging viral diseases - Ebola, Marburg, SARS, Hanta, Chikungunya, Zika, Chondipura; Antiviral chemotherapy and Viral vaccines; Nucleotide and nucleoside analogs, Reverse transcriptase inhibitor, protease inhibitor, fusion inhibitor etc., Interferons, Killed and attenuated vaccines.

Unit III

Fungal and protozoan infections

7 lectures

Types of Mycoses (with specific example of causative fungi) - Superficial, Cutaneous, Sub-cutaneous; Types of Mycoses (with specific example of causative fungi) - Endemic and Opportunistic; Mycotoxins and Antifungal chemotherapy - Mycetismus, Aflatoxins, classes of currently available drugs and new inhibitors in the pipeline; Protozoan diseases - Giardiasis, Amoebiasis; Leishmaniasis, African sleeping sickness; Malaria, Cryptosporidiosis; Infection by Helminths - Nematodes, Trematodes, Cestodes.

Unit IV

Sexually transmitted diseases and congenital infections

6 lectures

Syphilis and Gonococcal infections; AIDS and Lentiviral infection; Herpes infections; Chlamydial infections (*Chlamydia trachomatis*); Mycoplasma and Ureaplasma infection; Toxoplasmosis; Congenital viral infections - Cytomegalovirus, Varicella zoster, HBV, Enterovirus, Parvovirus B19 etc.

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Revised Biochem Curricula | 285



Unit V
Host-pathogen interaction
5 credits

Intracellular and extracellular pathogens, Principles of microbial pathogenesis, host damage, inflammatory responses, adaptation strategies of pathogen- impact of host and pathogen metabolism on immunity and pathogen survival; Chronic pathogens and mechanisms of persistence; Evasion mechanisms of pathogens; Bacterial - host interaction- *Mycobacterium tuberculosis*, *Borrelia burgdorferi*; Viruses - host interaction: HIV, Influenza; Protozoan - host interaction: *Plasmodium* spp., *Leishmania major*.



Recommended Textbooks and References:

1. KC Carroll, SA Morse, T Mietzner, S Miller. (2016) *Jurvetz, Melnick and Adelbergs's Medical Microbiology 27th edition*, McGraw Hill.
2. J Owen, J Punt and Sharon Stanford, (2012), *Kuby Immunology*, 7th edition, W. H. Freeman and Co.
3. IT Kudva, NA Cornick, PJ Plummer, Q Zhang, TL Nicholson, JP Bannantine and BH Bellare. *Virulence Mechanisms of Bacterial Pathogens*, (2016) 5th edition, ASM Press.
4. V Kumar, AK, Abbas and JC Aster, (2015), *Rabbits & Cotran Pathologic Basis of Disease*, 9th Edition, Elsevier.
5. K Murphy and K Weaver, (2016), *Janeway's Immunobiology*, 9th Edition, Garland Science.
6. AK Abbas, (2015), *Cellular and Molecular Immunology*, 8th Edition, Elsevier.
7. Anantharaman and Paniker, *Textbook of Microbiology*, 8th Edition.
8. Baveja CP, (2001) *Textbook of Microbiology*, 5th Ed., McGraw Hill Education.

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Course: Animal Biotechnology

Course Code: MBT 309T

Course Credit: 3

Unit I

Introduction to the balanced salt solutions and simple growth medium. Brief discussion on the chemical, physical and metabolic functions of different constituents of culture medium, Serum & protein free defined media and their applications.

Unit II

Primary and secondary cell culture, Development of cell lines, Biology and characterization of the cultured cells. Basic techniques of mammalian cell cultures in vitro.

Unit III

Maintenance of cell culture, Cell Passaging, Measuring parameters of growth, Measurement of viability and cytotoxicity.

Unit IV

Cell synchronization, Cell transformation, Apoptosis, Cryopreservation, Common cell culture contaminants.

Unit V

Applications of animal cell culture: cell culture based products, vaccines, Hybridoma technology and monoclonal antibodies, stem cells and their applications, Animal cloning, IVF technology, Organ, organotypic and histotypic cultures.

Suggested Readings

1. Culture of Animal Cells: Freshney
2. Animal Cell Culture: John RW Masters
3. Animal Cell Culture Techniques: Martin Clynes
4. Transgenic Animals: Generation and Use: Louis-Marie Houdebine

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